

# Intron evolution in Fungi

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Taylor Lab

# Evolutionary genomics

## Evolution & Organismal

Phenotype  
Population structure  
Ecological adaptation  
Niche changes  
Phylogeny

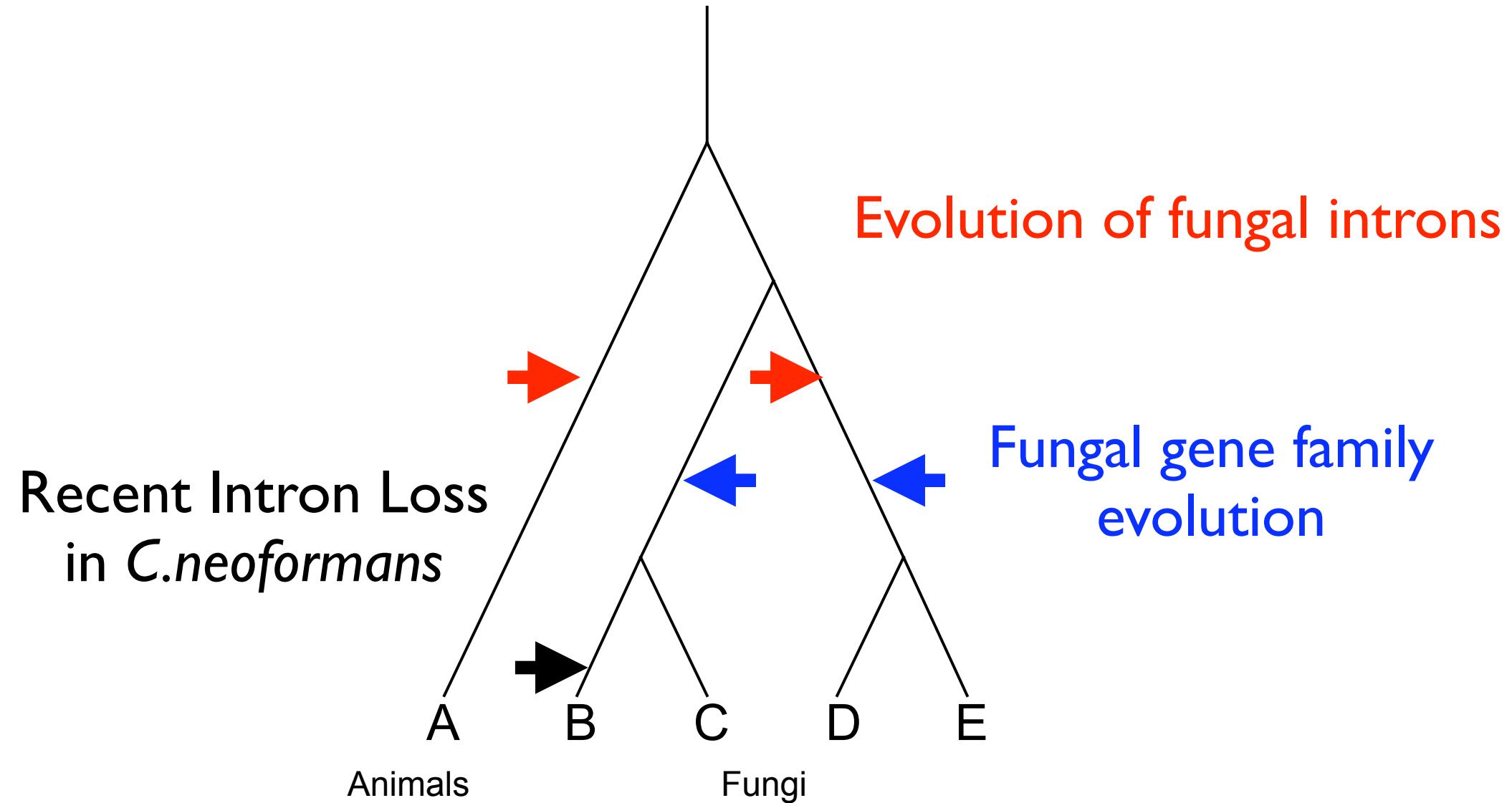
## Comparative Genomics

Molecular evolution  
Gene order  
Gene families  
Gene and genome structure  
Gene content  
Conserved elements  
Rates of molecular evolution

## Model Systems

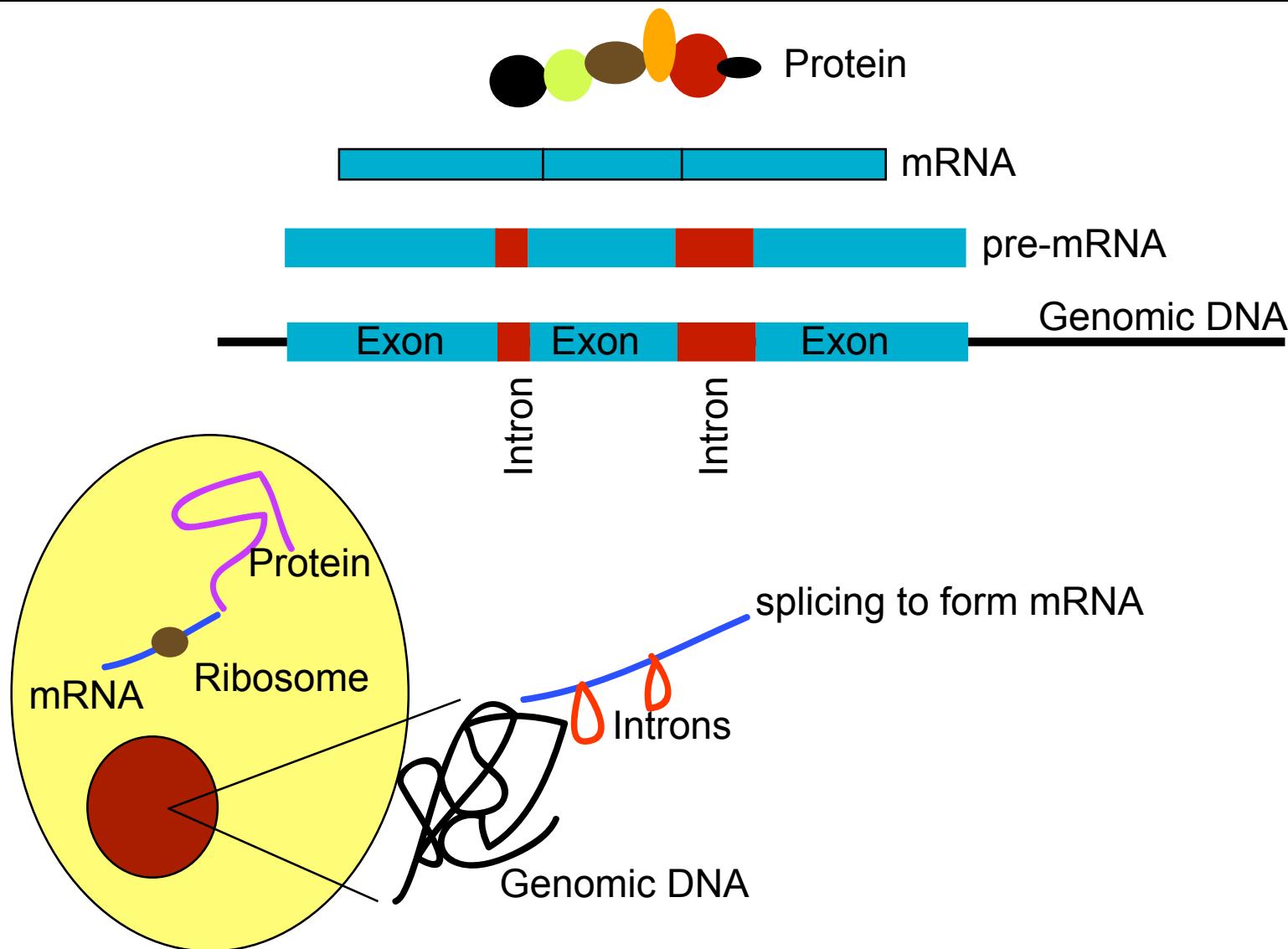
Genetic tools  
Gene function & expression  
Regulatory networks  
Pathways  
Molecular & cellular biology  
Disease models

# Fungal comparative genomics

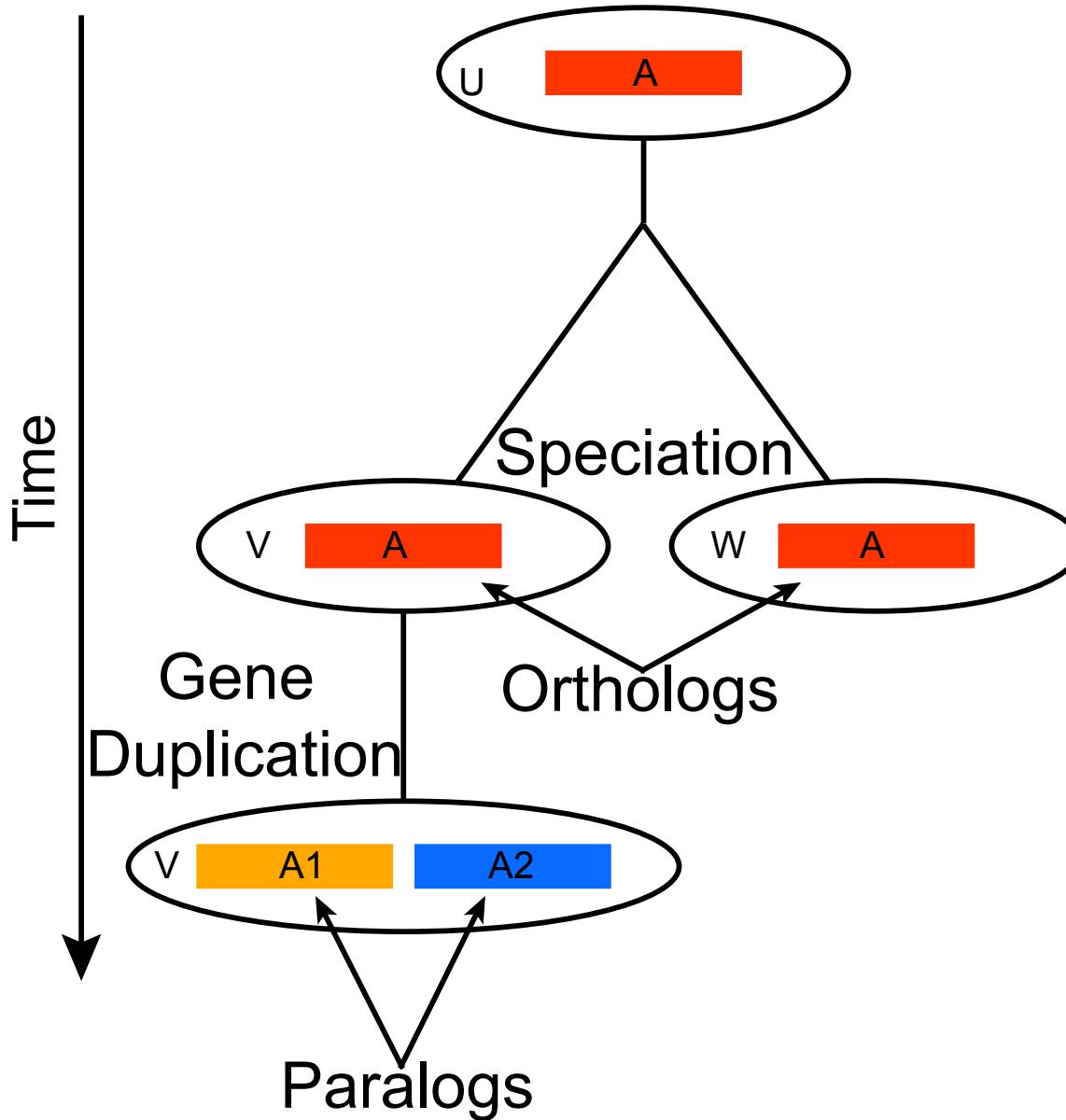


# Central dogma of eukaryotic biology

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# Orthologs and Paralogs

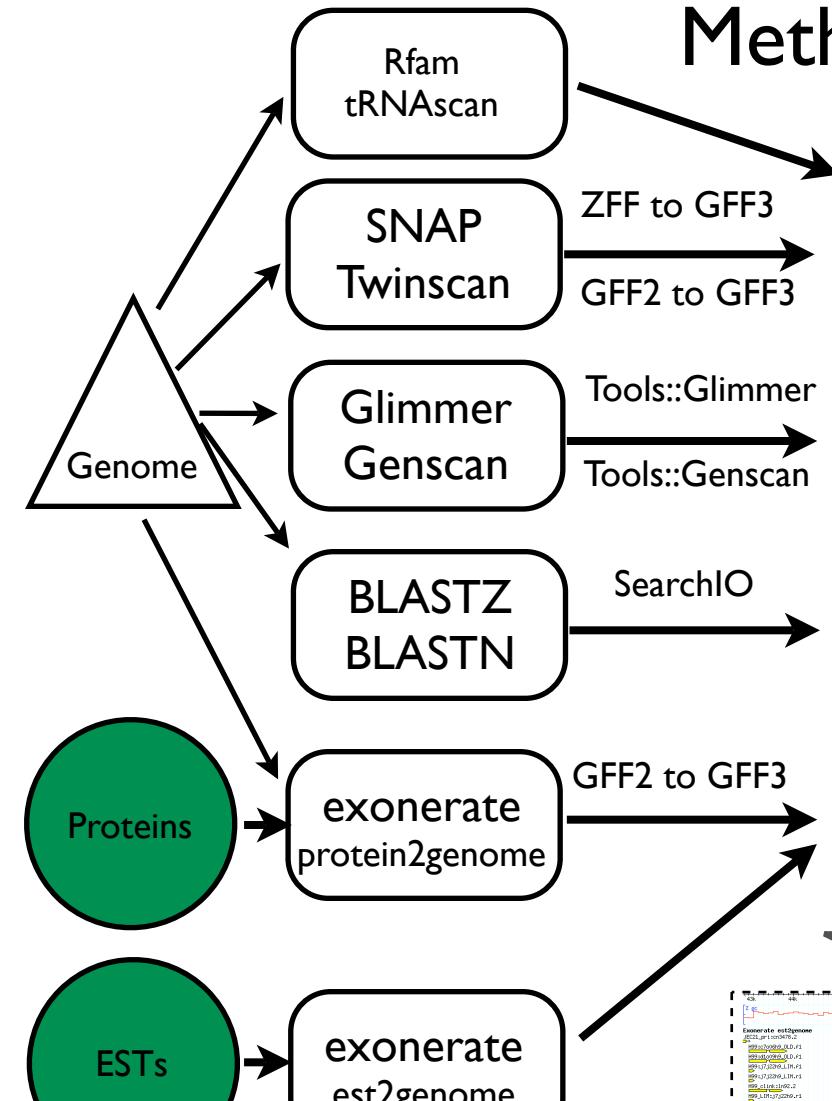


# Genome annotation

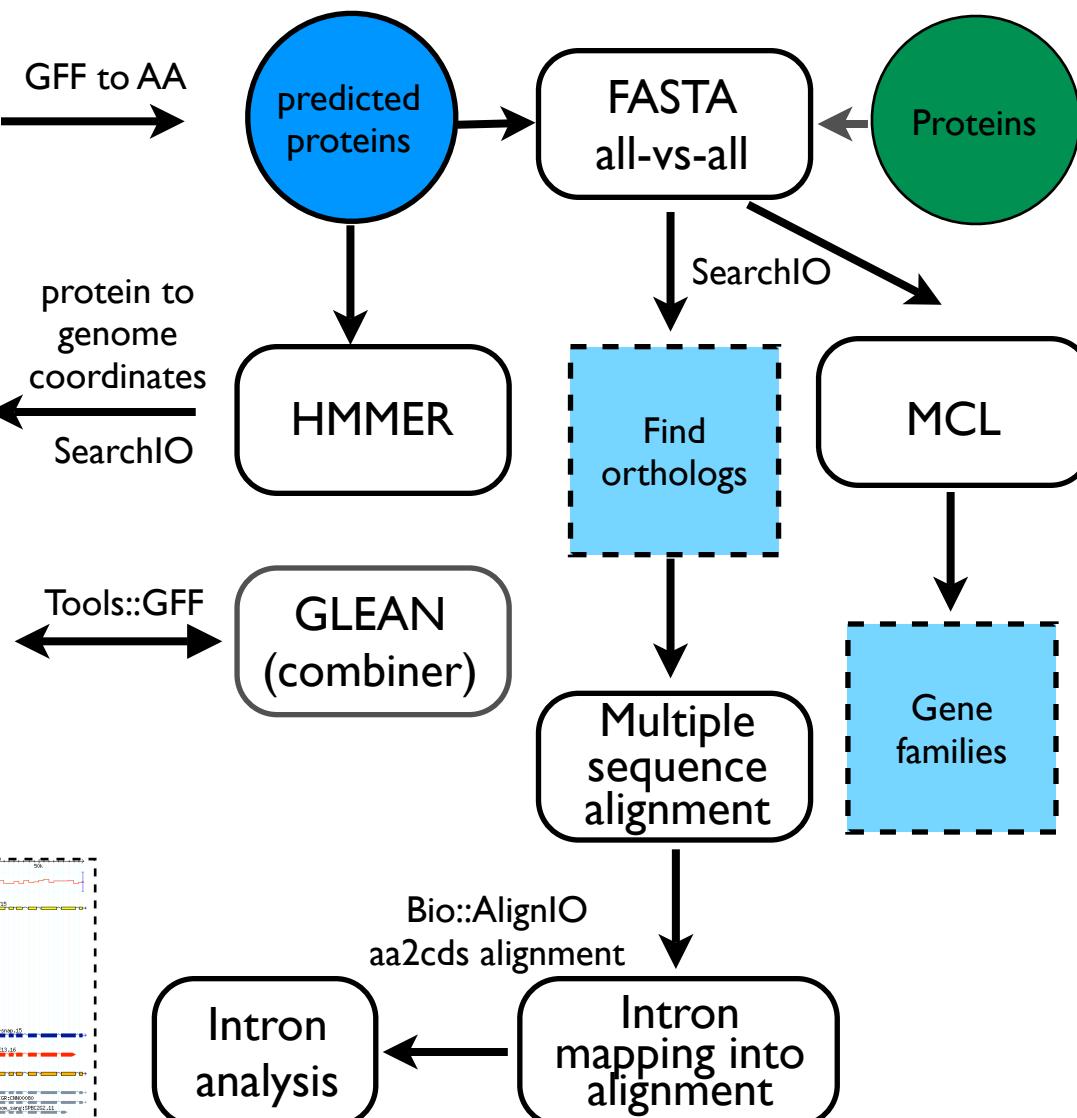
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- Many of the fungal genomes were only assembled genomic sequence.
- Automated annotation pipeline was built to generate to get systematic gene prediction.
- Several gene prediction programs were trained and results were combined to produce composite gene calls

# Methodology



# Analysis



<http://fun gal.genome.duke.edu>

# Evolution of gene structure

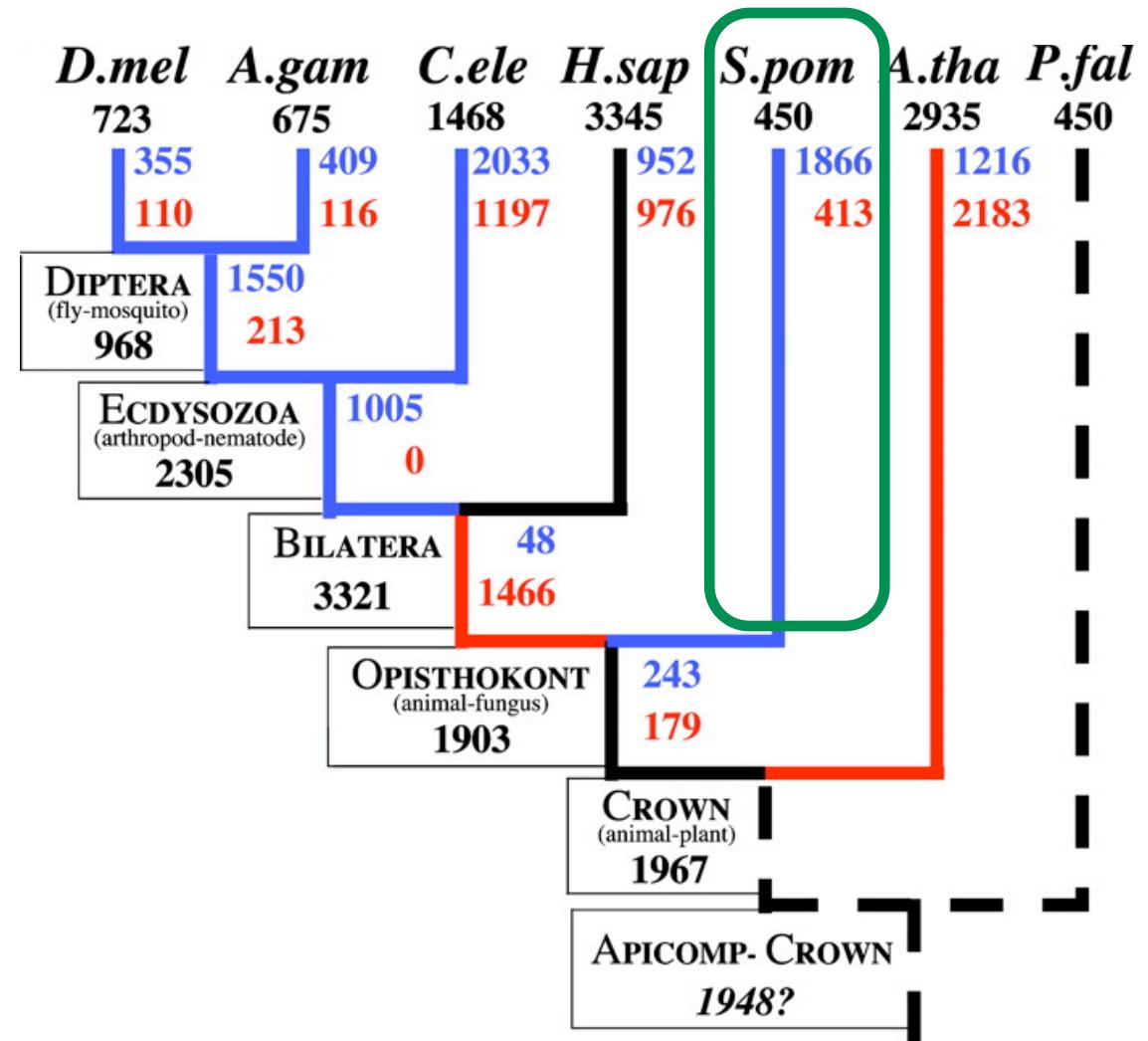
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- Present day introns
- Recent insertions?
  - Introns late hypothesis
  - Formed in eukaryotic ancestor?
  - Introns early hypothesis / exon theory of genes
- Mixture of two?

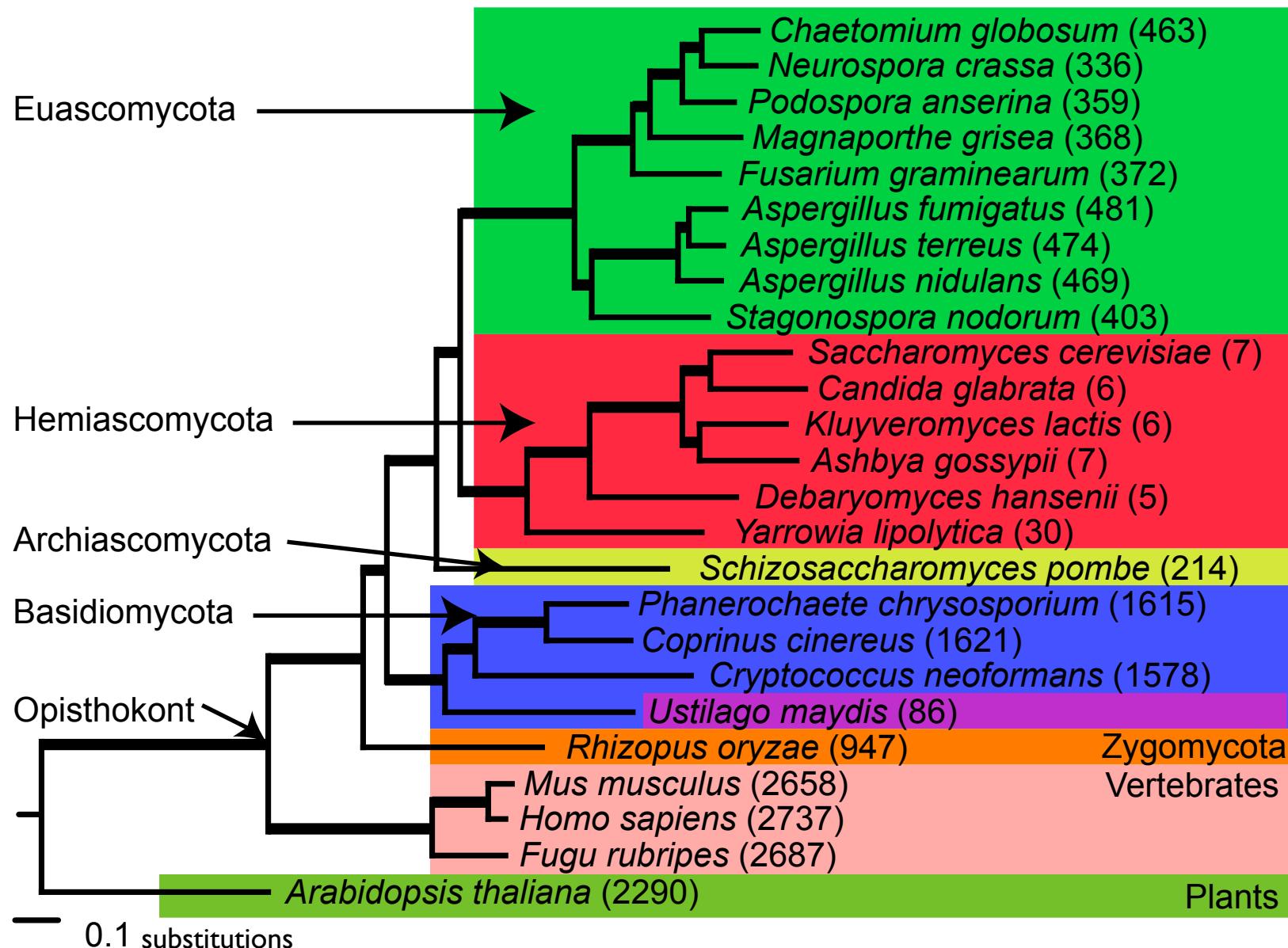
# Previous work on intron evolution

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- Rogozin et al. 2003
  - 7 genomes
  - 684 genes, 7236 positions
- Other methods
  - Roy and Gilbert. 2005
  - Csűrös. 2005
  - Nguyen et al. 2006

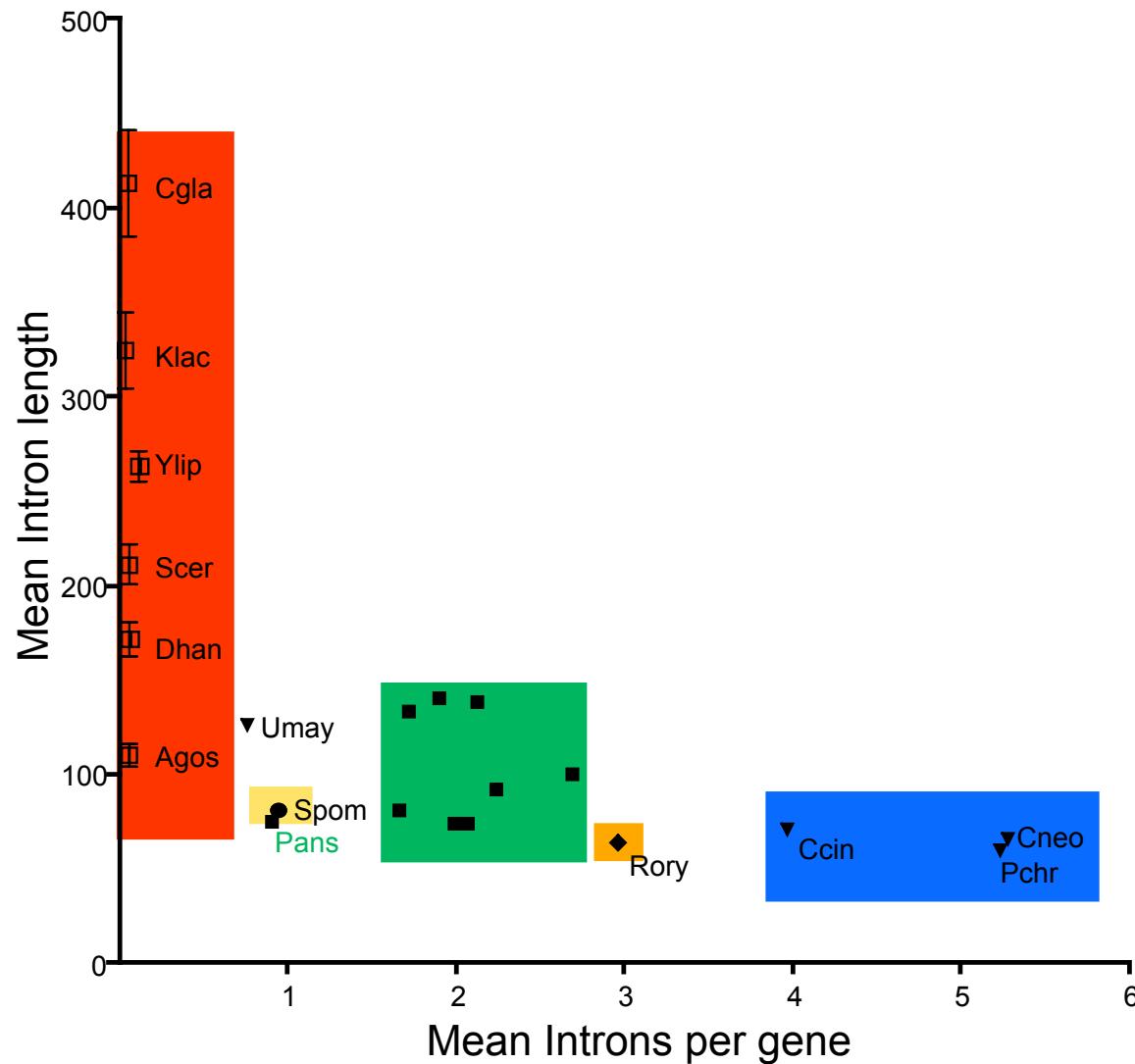


# Calculating intron densities across a phylogeny



# Intron frequency varies among the fungi

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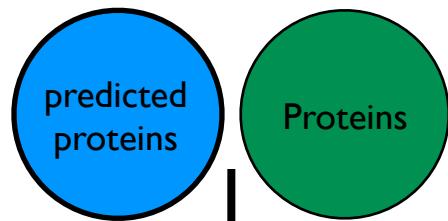
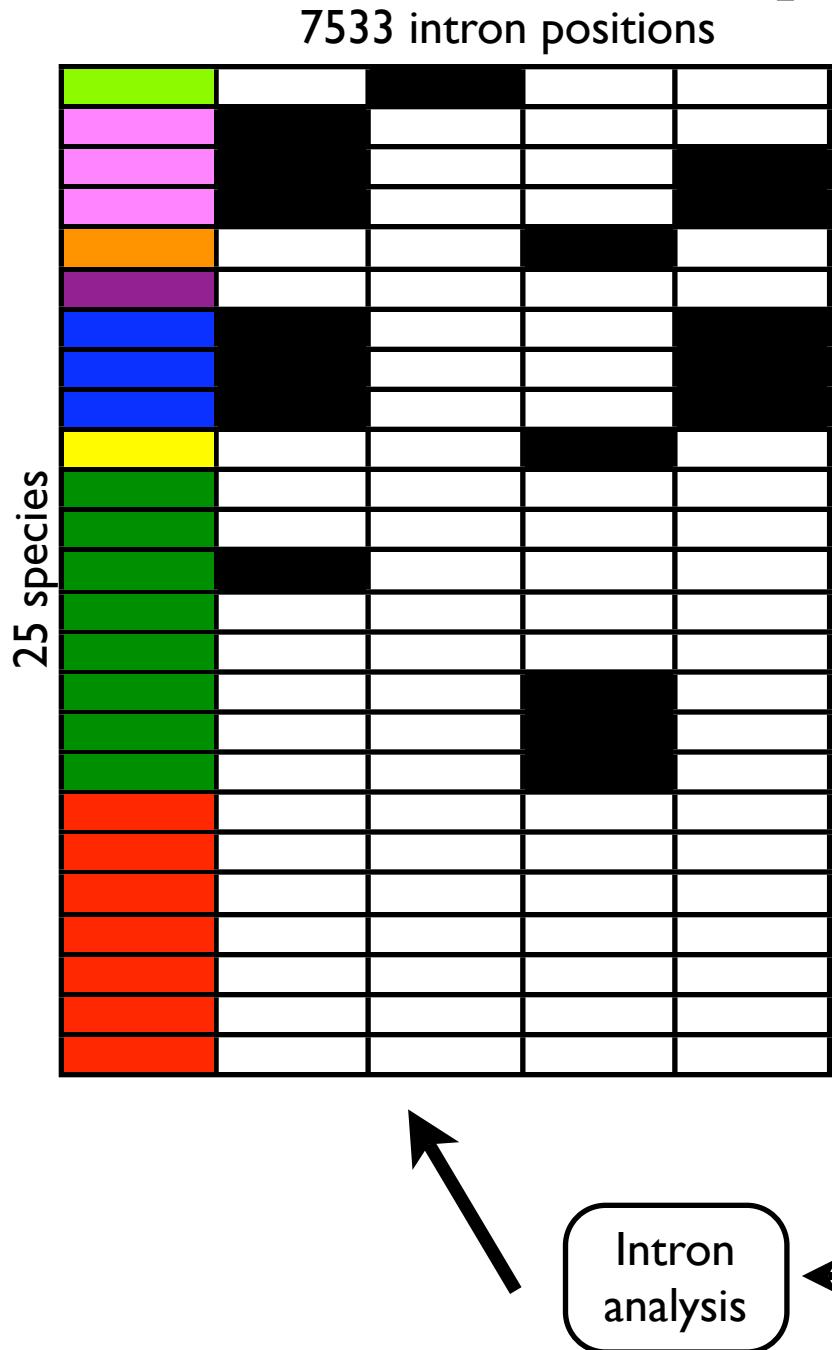


# Analysis of whole genomes

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- 25 entire genomes
  - 21 fungi, 3 vertebrates, 1 plant
- Largest dataset ever assembled for intron analysis
- 1160 orthologous genes
- 7533 intron positions
- 4.15 Mb coding sequence (CDS) per genome

# Analysis Methods

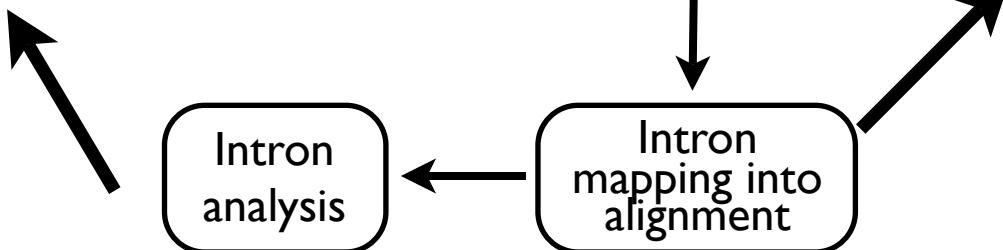


FASTA  
all-vs-all

Find  
orthologs

Multiple  
sequence  
alignment

Intron  
mapping into  
alignment

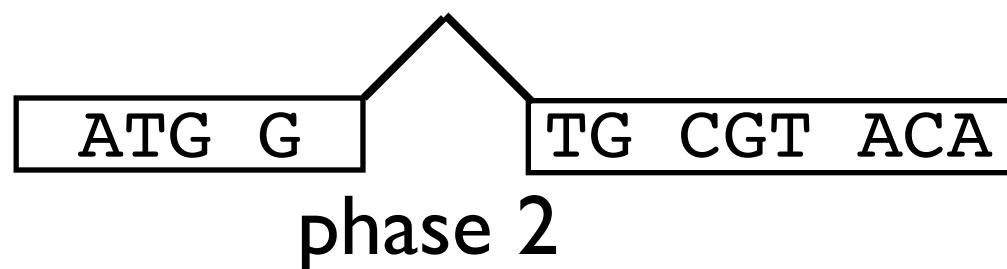
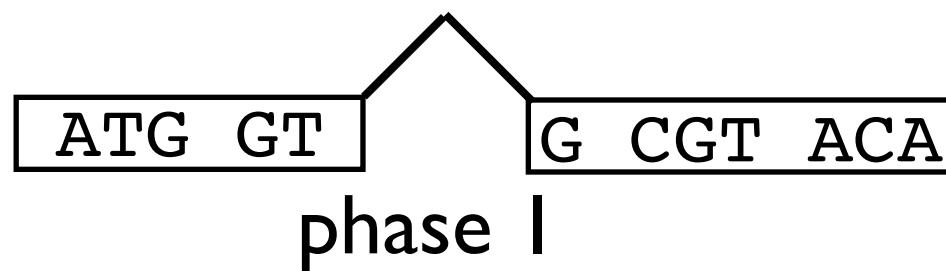
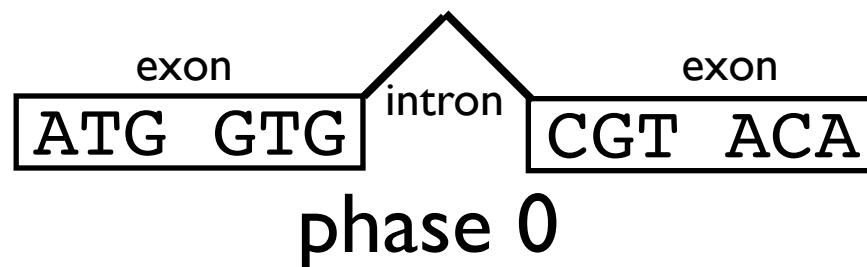


<i>A.thaliana</i>	TWLPEQVA-FIQS1MG-NDKANS-YW
Fugu	QWTQEQQVQ <b>0</b> SVQE-MG-NAKAKR-LY
Human	QWTQEQQ <b>0</b> CMQE-MG-NGKANR-LY
Mouse	QWTQEQQ <b>0</b> CMQE-MG-NGKANR-LY
<i>R.orzae</i>	TWVQDQVE-NMVR-WG-NERANK <b>2</b> YW
<i>U.maydis</i>	TWTREQVD-RMKE-VG-NLKSNR-KY
<i>C.neoformans</i>	IWTPEQME <b>0</b> SIQK-WG-NKRANM-YW
<i>C.cinereus</i>	VWTPEQME <b>0</b> SIQK-WG-NRRANL-YW
<i>P.chrysosporium</i>	VWTVEQMN <b>0</b> SIQK-WG-NKRANI-YW
<i>S.pombe</i>	SWTDEQTE-NMTR-WG-NERANL <b>2</b> YW
<i>F.graminearum</i>	SWTDEQLQ-SVLK-WG-NARANK-YW
<i>N.crassa</i>	AWTDEQLQ-SVLN-WG-NARANK-YW
<i>C.globosum</i>	SWTNEQVD <b>0</b> NMRK-VG-NVVSNK-LY
<i>P.anserina</i>	SWTDEQLQ-SVLN-WG-NARANK-YW
<i>M.grisea</i>	SWTDEQLQ-SVLS-WG-NARANK-YW
<i>S.nodorum</i>	TWTDEQLQ-SVLK-WG-NARANK-YW
<i>A.nidulans</i>	AWTDEQLQ-SVVR-WG-NARANK <b>2</b> YW
<i>A.fumigatus</i>	SWTDEQLQ-SVIK-WG-NARANK <b>2</b> YW
<i>A.terreus</i>	SWTDEQLQ-SVLR-WG-NARANK <b>2</b> YW
<i>Y.lipolytica</i>	AWTEEQLA-SMMK-WG-NTRCNM-FW
<i>D.hansenii</i>	AWTDEQVE-LMIK-WG-NEKCNI-YW
<i>K.lactis</i>	TWTEEHLE-AVLE-FGNKKFNE-YY
<i>A.gossypii</i>	TWKEEHLQ-QVVR-FGNNQQANK-VI
<i>C.glabrata</i>	TWKEEHLK-ELVQ-MRNNVNANR-VI
<i>S.cerevisiae</i>	TWKEEHLV-KLIQ-FKNNLRANS-YI

\* : : : \* : :

# Intron phase

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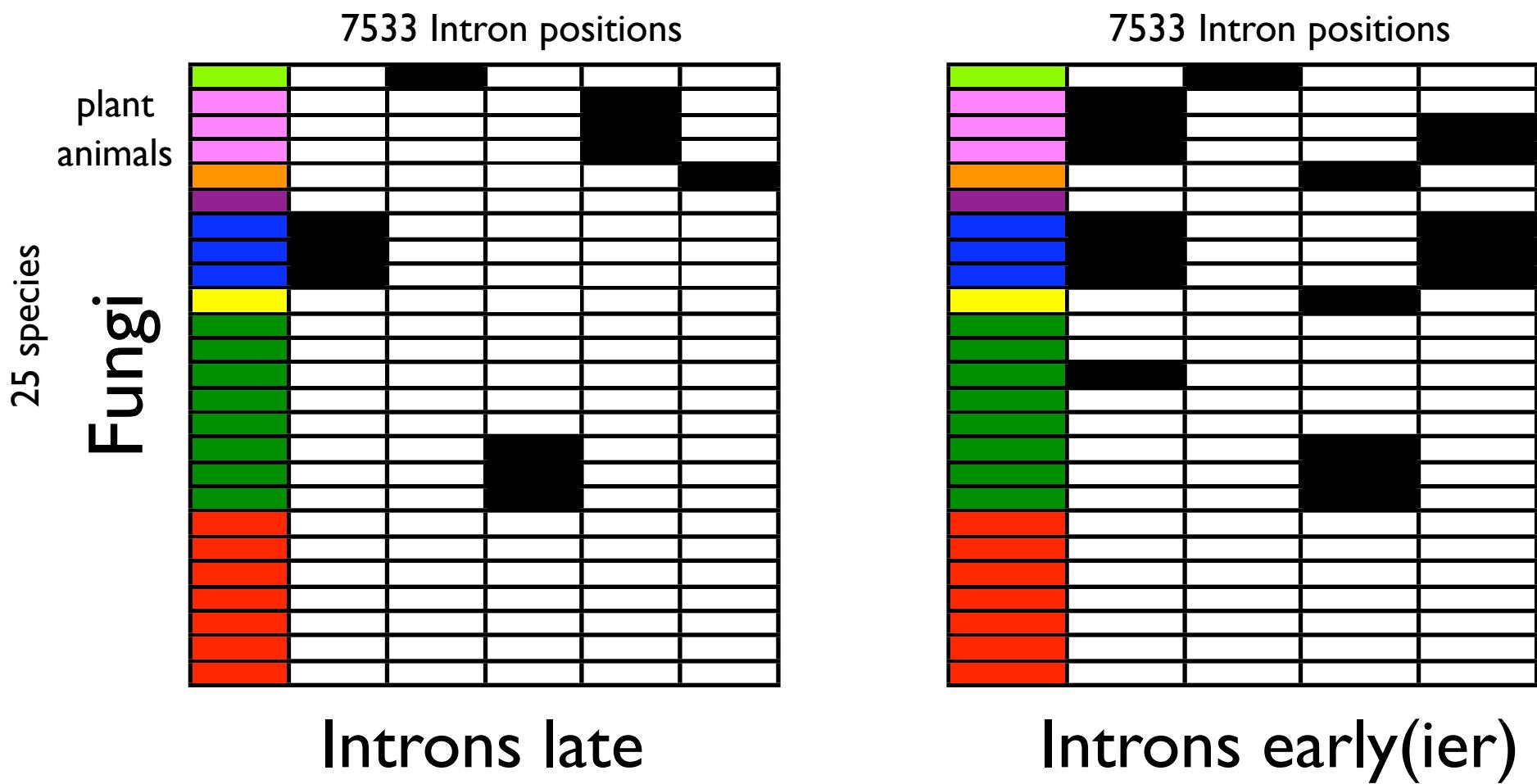


# Conserved intron positions

	TWLP <span style="background-color: black; color: white;">EQVA</span> -FIQS <span style="background-color: red; color: black;">1</span> MG-NDKANS-YWEA-----ELPP-----NYD-----RV-GIENFIRAK2Y-----EEKRWV--
A. thaliana	TWLP <span style="background-color: black; color: white;">EQVA</span> -FIQS <span style="background-color: red; color: black;">1</span> MG-NDKANS-YWEA-----ELPP-----NYD-----RV-GIENFIRAK2Y-----EEKRWV--
Fugu	QWTQE <span style="background-color: black; color: white;">QVQ</span> 0SVQE-MG-NAKAKR-LYE-----FLPK-----CFQRPETDQ-SAEIFIRDK-Y-----DKKKYMDK
Human	QWTQE <span style="background-color: black; color: white;">QIQ</span> 0CMQE-MG-NGKANR-LYE-----YLPE-----TFRRPQIDP <span style="background-color: red; color: black;">2</span> AVEGFIRDK-Y-----EKKKYMDR
Mouse	QWTQE <span style="background-color: black; color: white;">QIQ</span> 0CMQE-MG-NGKANR-LYE-----YLPE-----TFRRPQIDP <span style="background-color: red; color: black;">2</span> AVEGFIRDK-Y-----EKKKYMDR
R. oryzae	TWVQDQVE-NMVR-WG-NERANK2YWEA-----NL-----GDRKPS-ES-NMEMWIRAK-Y-----EQKRWA--
U. maydis	TWTREQVD-RMKE-VG-NLKSNR-KYNPDEMNRNPPT-----NMEESERDS-ELEKYIIRRK-Y-----EFRRFV--
C. neoformans	IWTPE <span style="background-color: black; color: white;">QME</span> 0SIQK-WG-NKRAM-NYWER-----HLKA-----GHI-PS-DH <span style="background-color: red; color: black;">2</span> KIESFIRSK-Y-----ETRRWA--
C. cinereus	VWTPE <span style="background-color: black; color: white;">QME</span> 0SIQK-WG-NRRANL-YWEA-----HLKP-----GHN-PP-EH <span style="background-color: red; color: black;">2</span> KMESFVRSK-Y-----ESRRWA--
P. chrysosporium	VWTVE <span style="background-color: black; color: white;">QMN</span> 0SIQK-WG-NKRANI-YWEA-----HLKA-----GHI-PP-DH <span style="background-color: red; color: black;">2</span> KMESFIRSK-Y-----ESKRWA--
S. pombe	SWTDEQTE-NMTR-WG-NERANL <span style="background-color: red; color: black;">2</span> YWEA-----KLAG-----GHV-PS-DS <span style="background-color: red; color: black;">2</span> KIATFIKTK-Y-----EFKKWV--
F. graminearum	SWTDEQLQ-SVIK-WG-NARANK-YWEA-----KLAA-----GHA-PS-EA-KIENFIRTK-Y-----ELKRWV--
N. crassa	AWTDEQLQ-SVLN-WG-NARANK-YWEA-----KLAQ-----GHV-PS-ES-KIENFIRTK-Y-----ELKRWV--
C. globosum	SWTNE <span style="background-color: black; color: white;">QVD</span> 0NMRK-VG-NVVSNK-LYNPDN-----KNPPVPIDADEA-----DG-AMERFIRQK-YIARTLSIGKRRPGGD
P. anserina	SWTDEQLQ-SVLN-WG-NARANK-YWEA-----KLAP-----GHV-PS-EA-KIENFIRTK-Y-----ELKRWV--
M. grisea	SWTDEQLQ-SVLS-WG-NARANK-YWES-----KLAA-----GHA-PS-EA-KIENFIRTK-Y-----ELKRWV--
S. nodorum	TWTDEQLQ-SVIK-WG-NARANK-YWEA-----KLAP-----GHV-PS-EA-KIENFIRTK-Y-----ESKRWT--
A. nidulans	AWTDEQLQ-SVVR-WG-NARANK <span style="background-color: red; color: black;">2</span> YWEA-----KLAP-----GHV-PP-EA <span style="background-color: red; color: black;">2</span> KIENFIRTK-Y-----ESKRWV--
A. fumigatus	SWTDEQLQ-SVIK-WG-NARANK <span style="background-color: red; color: black;">2</span> YWEA-----KLAP-----GHV-PS-EA <span style="background-color: red; color: black;">2</span> KIENFIRTK-Y-----ESKRWV--
A. terreus	SWTDEQLQ-SVLR-WG-NARANK <span style="background-color: red; color: black;">2</span> YWEA-----KLAP-----GHV-PS-EA <span style="background-color: red; color: black;">2</span> KIENFIRTK-Y-----ESKRWV--
Y. lipolytica	AWTEEQLA-SMMK-WG-NTRCNM-FWEA-----KLPK-----GHV-PD-DN-KIENFIRTK-Y-----DMKKWA--
D. hansenii	AWTDEQVE-LMIK-WG-NEKCNI-YWES-----KLPD-----GYV-PD-QL-KIDNFIRTK-Y-----DLKKWV--
K. lactis	TWTEEHLE-AVLE-FGNNKKFNE-YYEN-----KLGG-----GTYVPD-QS-KIGQFIRTK-Y-----ELKKWV--
A. gossypii	TWKEEHLQ-QVVR-FGNNQQANK-VFEG-----RLGG-----GSYVPD-QS-KMGQFIKTK-Y-----EVRKWY--
C. glabrata	TWKEEHLK-ELVQ-MRNNVNANR-VYEA-----KLPDSSKFNGKSLGNDIN-LLQEFIRQK-Y-----ERKRWL--
S. cerevisiae	TWKEEHLV-KLIQ-FKNNLRANS-YYEATL-ADELKQ-----RKI-TD-TS-SLQNFIFKNK-Y-----EYKKWI--

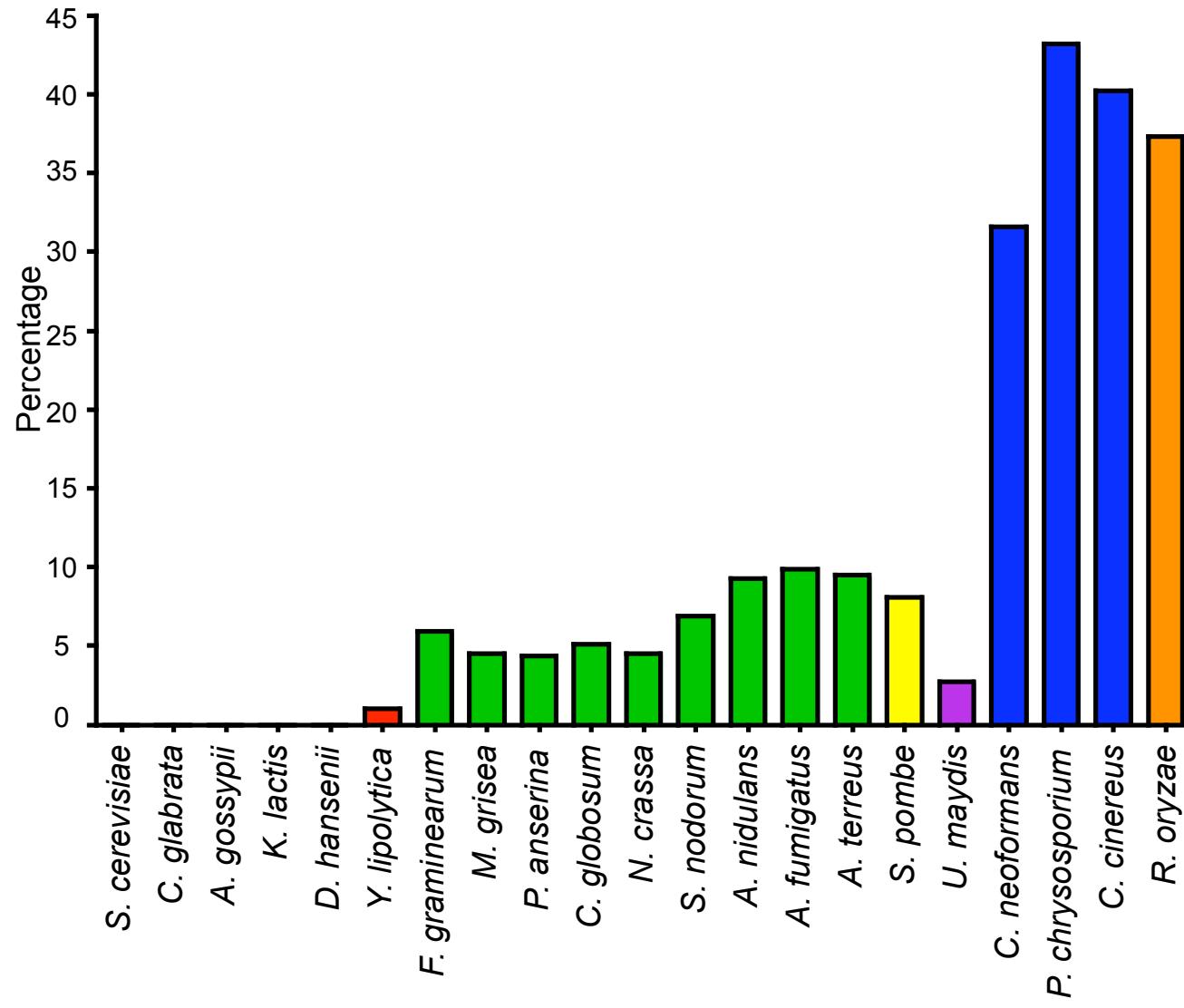
# Patterns of conservation

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# Intron positions shared with animals or plants

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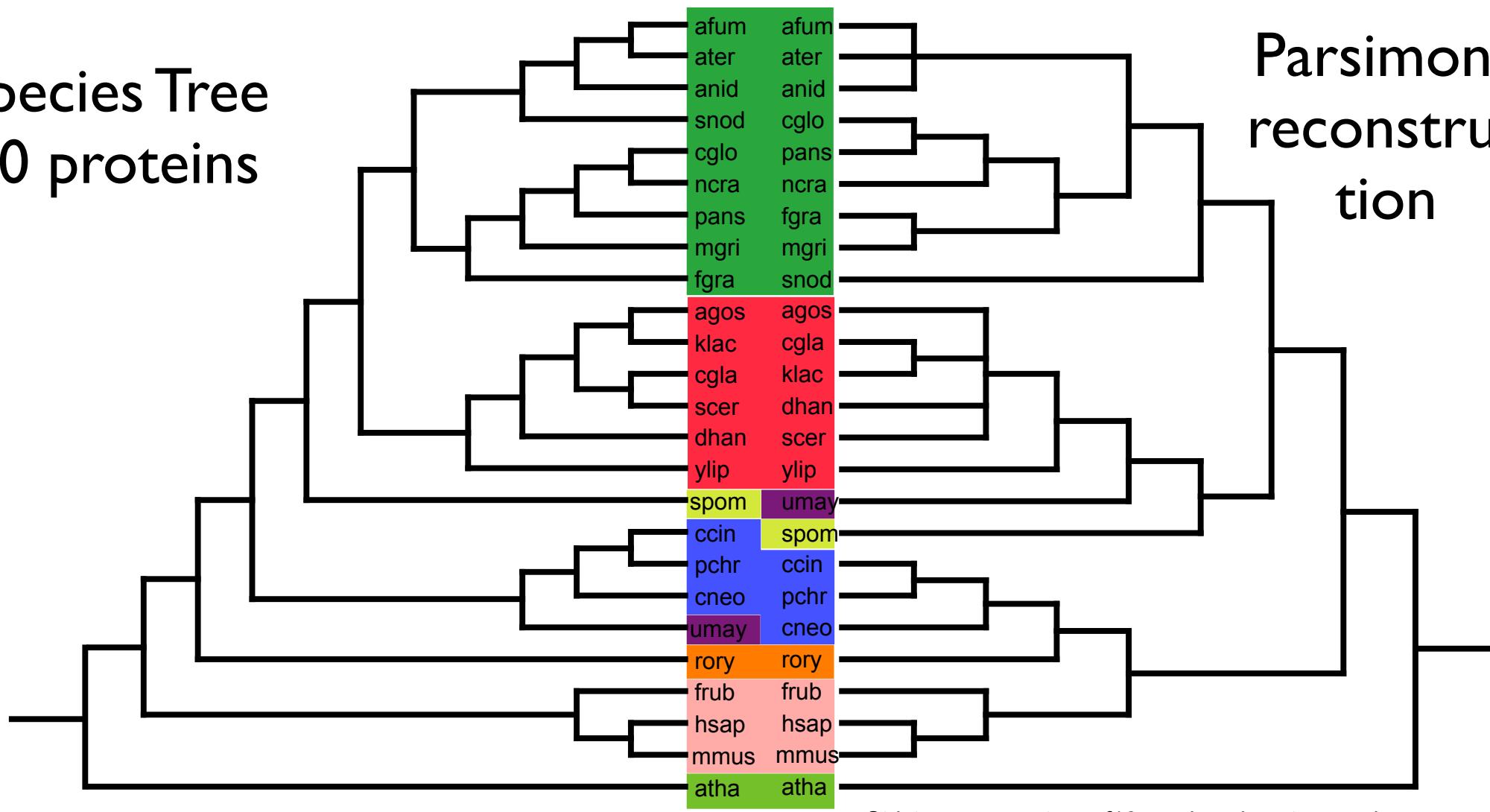


# Phylogenetic signal in intron positions

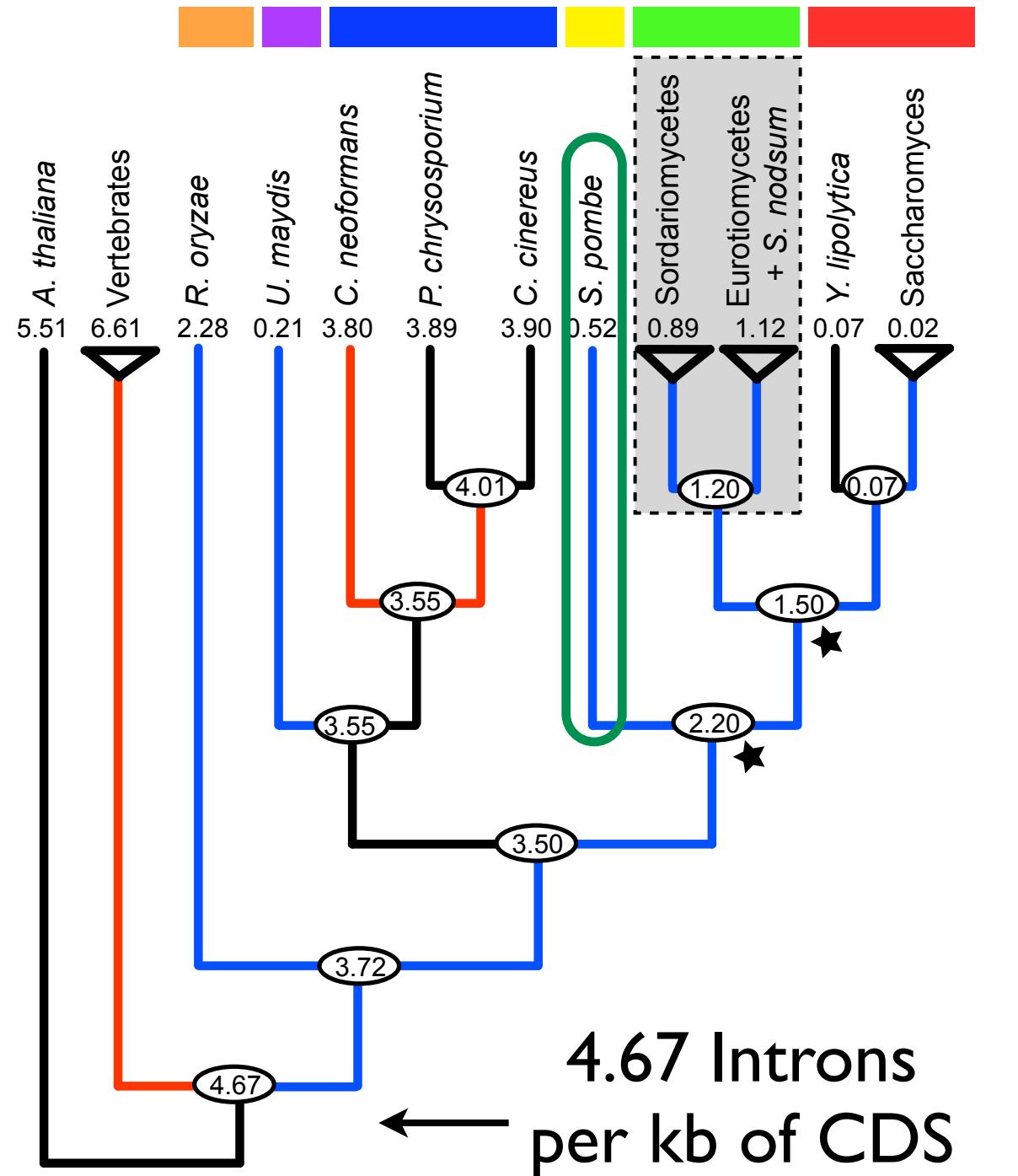
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Species Tree  
30 proteins

Parsimony  
reconstruction



# Reconstruction of ancestral intron densities



# Conclusions

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- Early eukaryotic crown genes were complex
  - Ancestor had 70% of the introns in vertebrates
  - More introns than previously reported
- Intron loss has dominated among the fungi
  - Hemiascomycota experienced loss
- Sampling can bias interpretations - all fungi are not equal.

# Mechanism of intron loss

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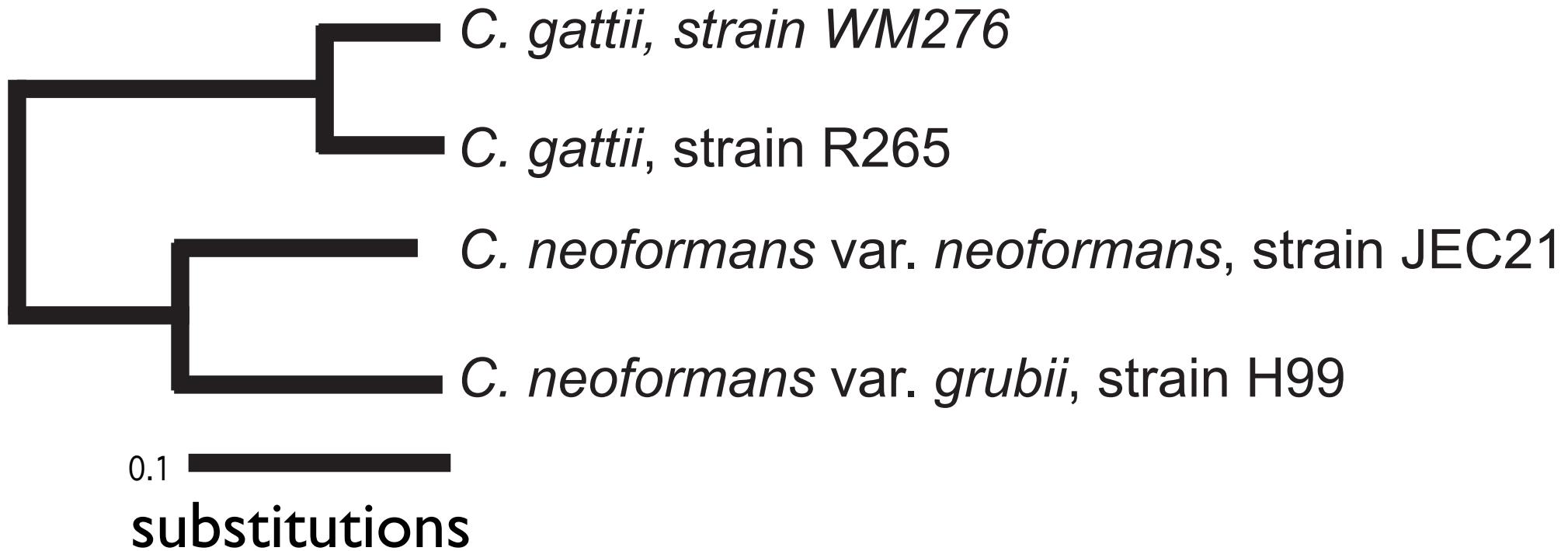
- *S. cerevisiae* and Hemiascomycota have undergone intron loss.
- How are introns lost from the genome?
  - Are they lost independently?
  - Are they lost many at a time?
- Molecular mechanism of loss

# Models of intron loss

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- All introns in *S. cerevisiae* are in 5' end of gene
- G. Fink proposed transcripts recombine with genome 3' -> 5' explaining 5' retention bias.
- In *S. cerevisiae* most intron loss events occurred too long ago so little evidence supporting any mechanism

# Sequenced *Cryptococcus* genomes

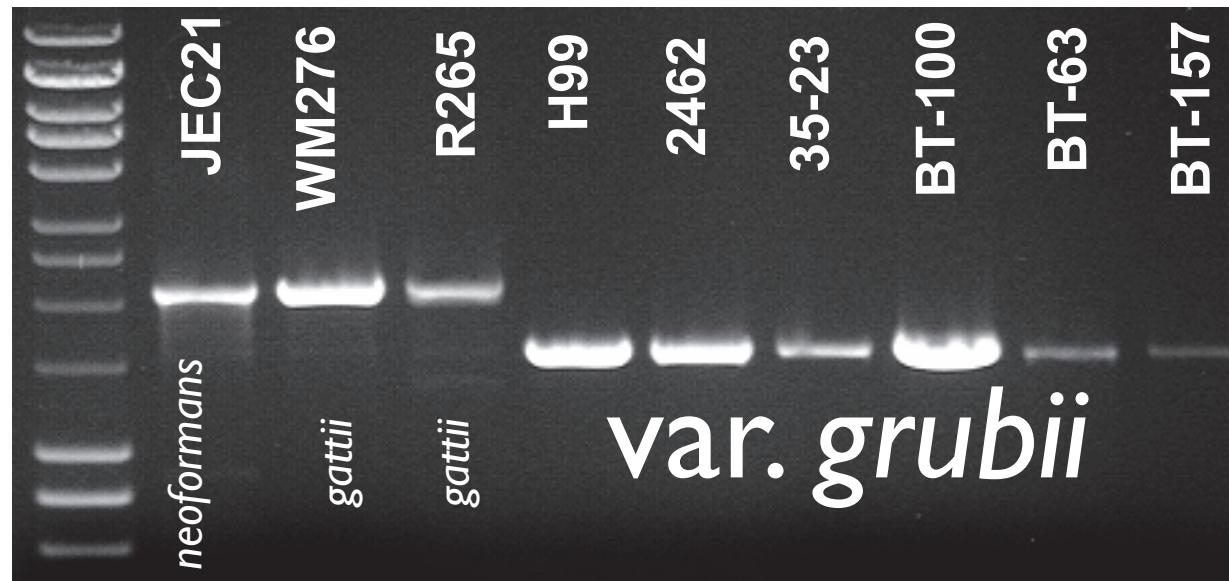


# Screen for intron changes

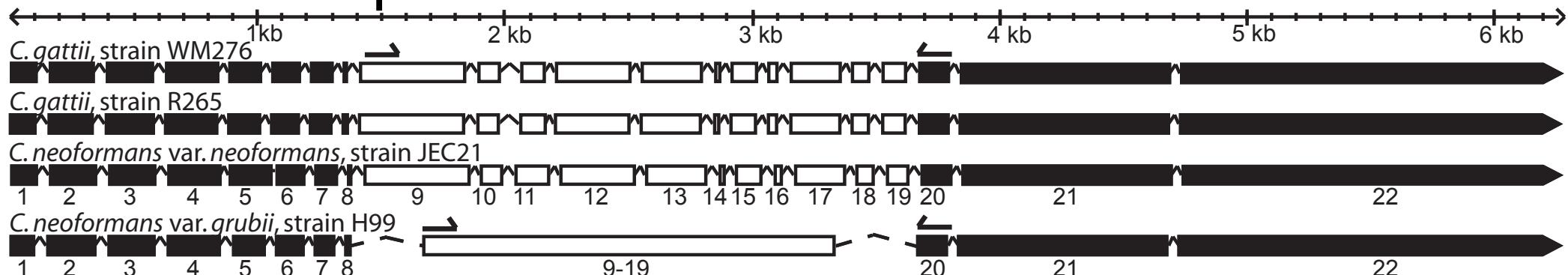
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- Annotate 3 *Cryptococcus* genomes (var. *grubii* and 2 var. *gattii* genomes)
- Identify and align 4-way orthologous genes
  - 5298 orthologous genes (out of ~6500)
- Identify intron position changes

# Intron loss in var. *grubii*



## CNI01550 - putative RNA helicase



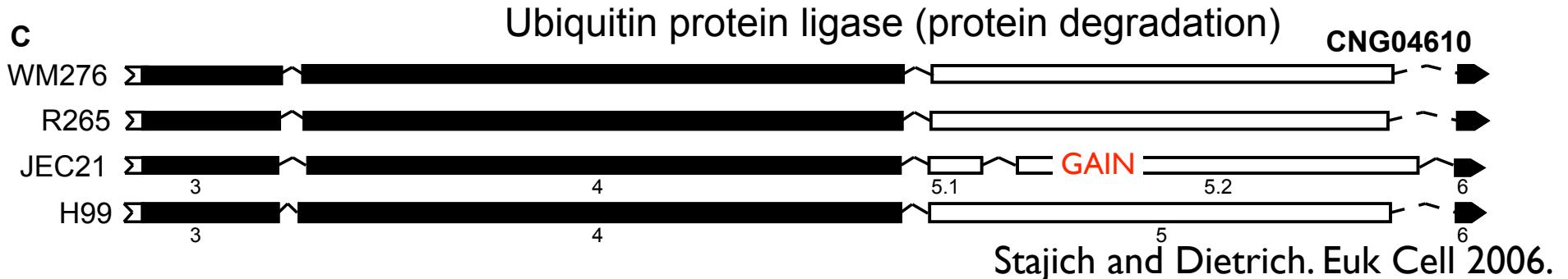
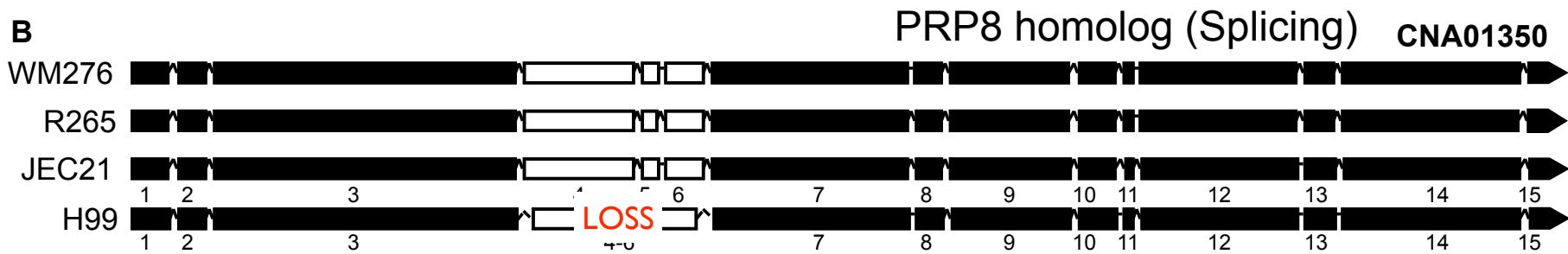
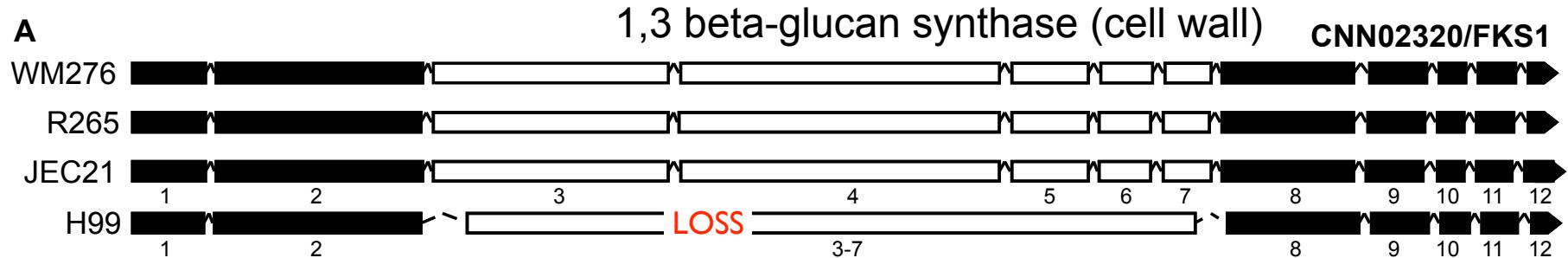
# Intron loss was a precise excision

R265	CGACAAGTACATAAAACTTTTGTGCCTGGCGCAAAGACTTCCATTGCTGACAGAAAACAGGTTGAA
WM276	AGACAAGTACATAAAACTTTTGTCTCTCCTCCAAACATTTCATTGCTGACAGAAAACAGGTTGAA
H99	AGACAA-  - GTTGA
JEC21_CDS	AGACAA-----GTTGAA
JEC21	AGACAAGTACATACTAGTCCTTG-----CTATCCAAAGACTTT-CATTGCTGACAGAAAACAGGTTGAA *****

R265	CGCTGCCGAATTATGTCGATGTTGGAGATTCTTGAGGTAAGCAACAGACTCGTAACAGCTTGTTCGGT
WM276	CGCTGCCGAACTATGTCGATGTTGGAGATTCTTGAGGTAAGCAACAGACTCGTAACAGCTTGTTCGGT
H99	CCCTGCCGAATTATGTCGACGTTGGAGATTCTTGAG-----
JEC21_CDS	CCCTGCCGAATTATGTCGATGTTGGAGATTCTTGAG-----
JEC21	CCCTGCCGAATTATGTCGATGTTGGAGATTCTTGAG <b>GTACGTCGCAA</b> ACTCGTAACAGCTTGTTCGATC *****

# Other examples of loss



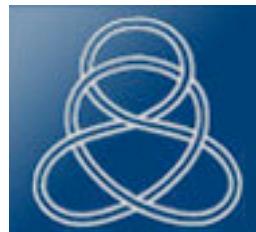
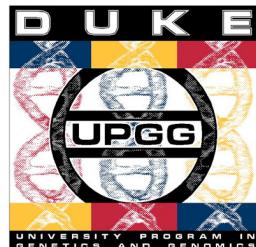
# Conclusions

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- Intron loss via homologous recombination with spliced transcript
  - Large losses are all adjacent introns
  - Precise deletion
- Loss biased towards the middle of gene not 3'

# Acknowledgements

Fred Dietrich  
Scott Roy



Sequencing centers  
Broad Institute  
Duke University  
Joint Genome Institute  
Génolevures  
Stanford University  
(NIH and NSF)