

# A Comparative Genomic Investigation of Fungal Genome Evolution

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# Evolutionary genomics

## Evolutionary & Organismal Biology

Phylogeny  
Population genetics and structure  
Phenotype  
Ecological adaptations

## Comparative Genomics

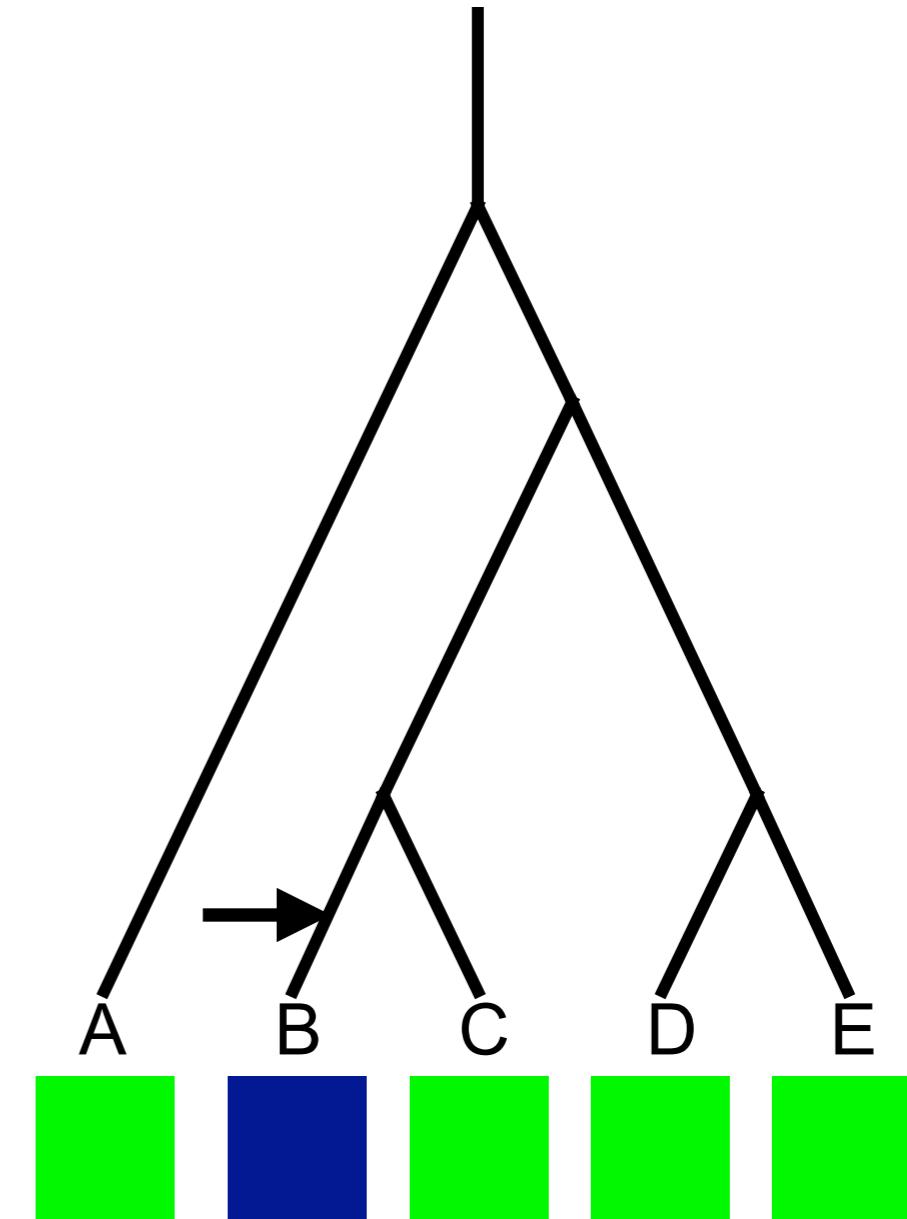
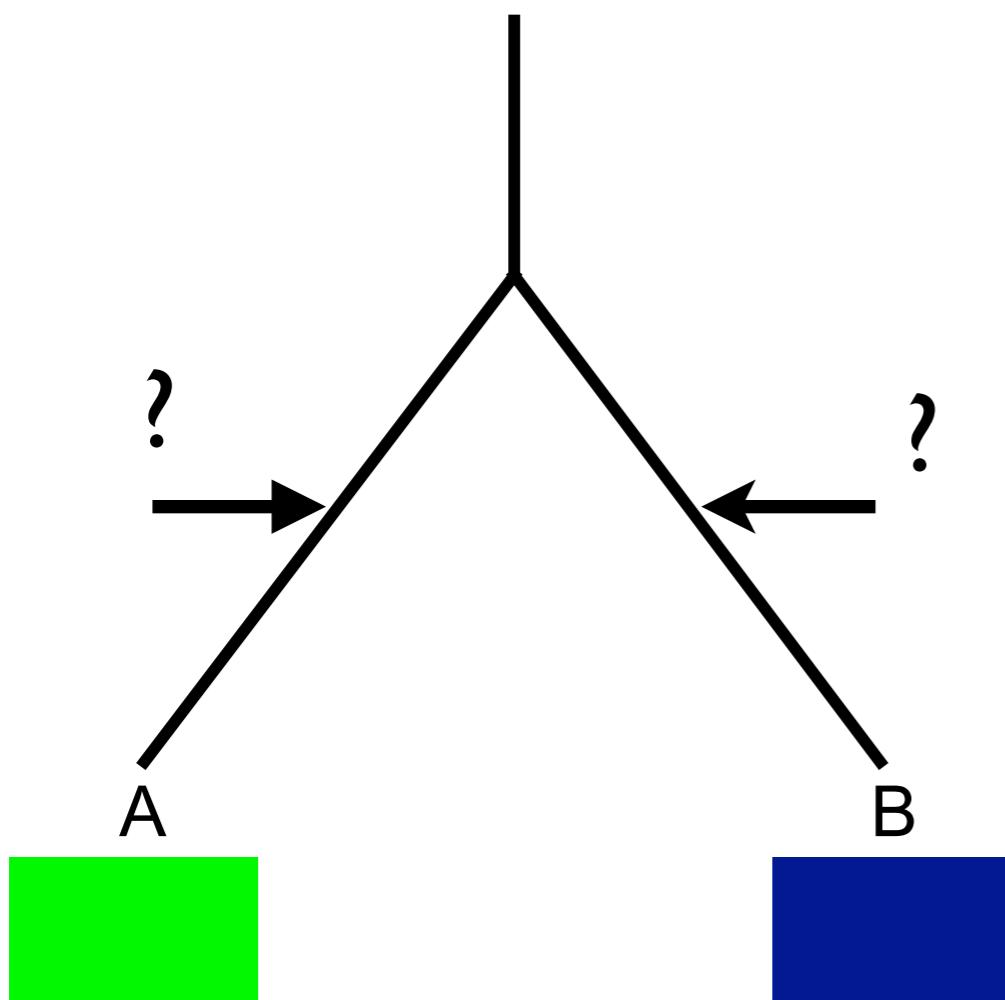
Molecular evolution  
Gene order  
Gene families  
Gene and genome structure  
Gene content  
Conserved elements  
Rates of molecular evolution  
Gene function inference

## Model Systems

Genetic tools  
Gene function & expression  
Regulatory networks  
Pathways  
Molecular & cellular biology  
Disease models

# Power of the comparative approach

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# Industrial uses of fungi

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- Bread, beer, wine - *Saccharomyces cerevisiae*
- Sake and soy sauce - *Aspergillus oryzae*
- Dairy - *Penicillium roqueforti*, *Kluyveromyces lactis*
- Citric acid - *Aspergillus niger*
- Riboflavin - *Ashbya gossypii*
- Stonewashed jeans - *Trichoderma reesei*
- Penicillin antibiotic - *Penicillium notatum*
- Button Mushrooms - *Agaricus bisporus*

# Agricultural impact of fungi

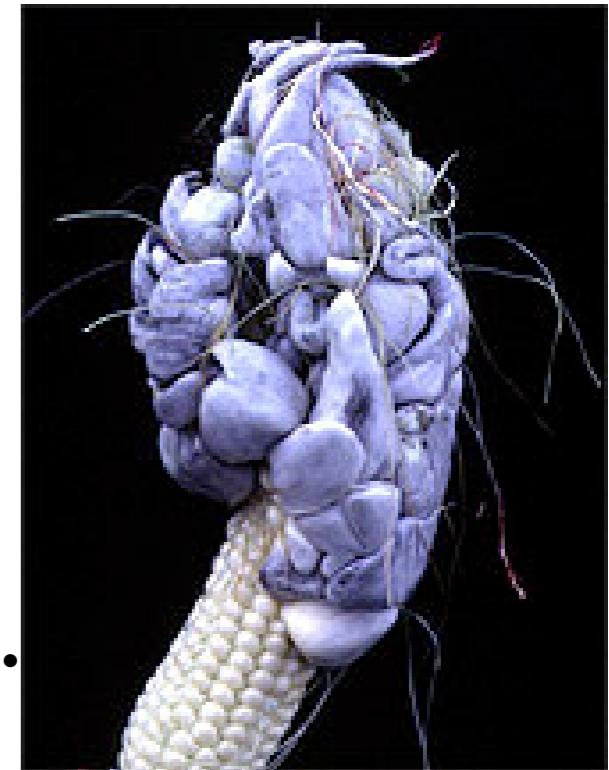
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causes of plant disease is caused by



deposit mycotoxins - e.g. ergot



beneficial fungi provide nutrient  
and nitrogen fixation

# Impact of fungi on human health

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- Mostly immunocompromised individuals are at risk of life-threatening infections
- Primary pathogens
  - *Histoplasma, Coccidioides, Cryptococcus gattii*
- Opportunistic pathogens
  - *Candida albicans, Aspergillus fumigatus, Cryptococcus neoformans, Rhizopus oryzae*

# Fungi as genetic models

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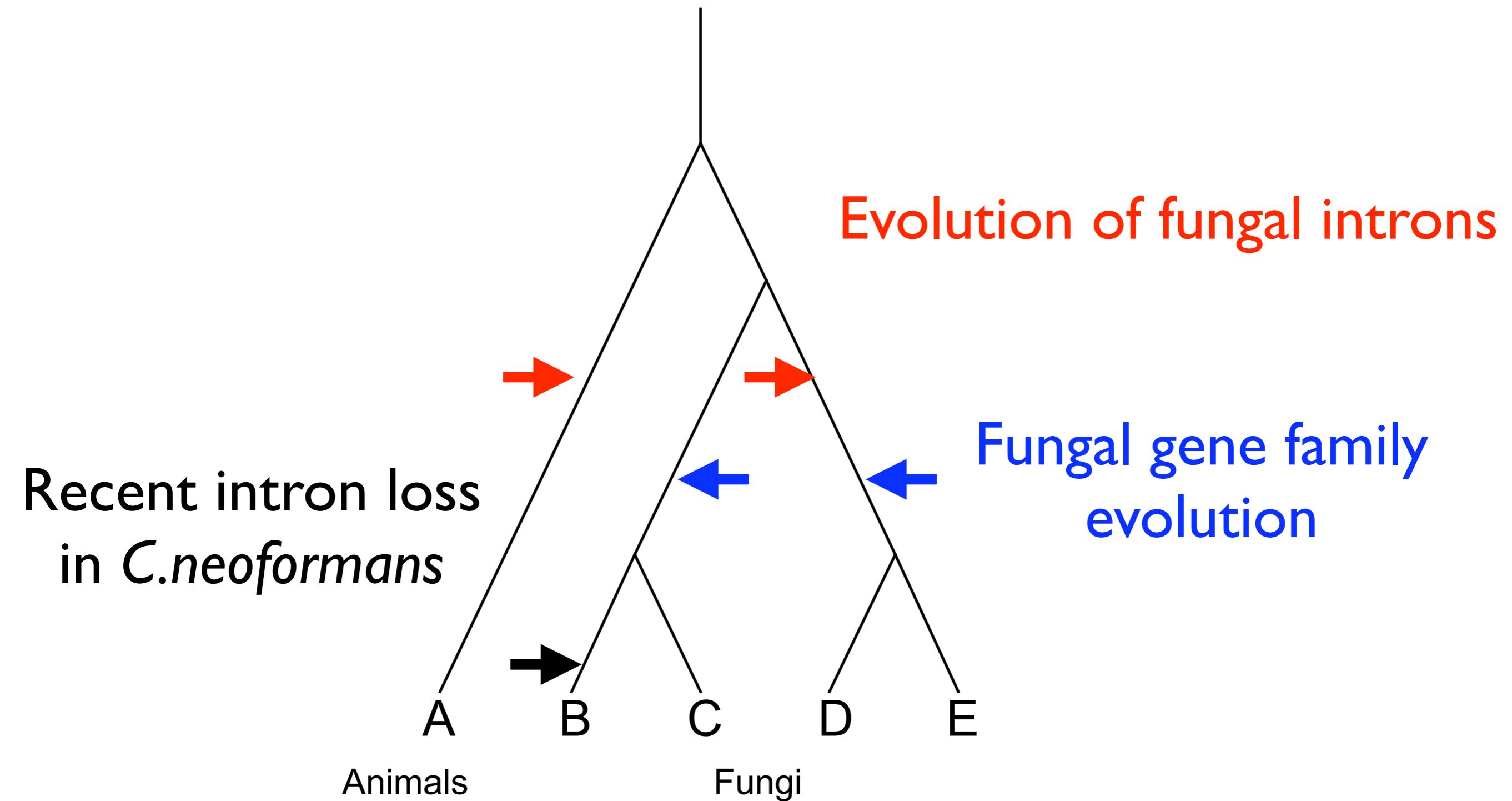
- Beadle and Tatum (1941) - one gene, one enzyme hypothesis in *Neurospora crassa*
- Cell cycle, cell model - *Saccharomyces cerevisiae* and *Schizosaccharomyces pombe*
- Molecular biology tools to investigate phenotype-genotype
- Evolutionary models

# Fungal genomes

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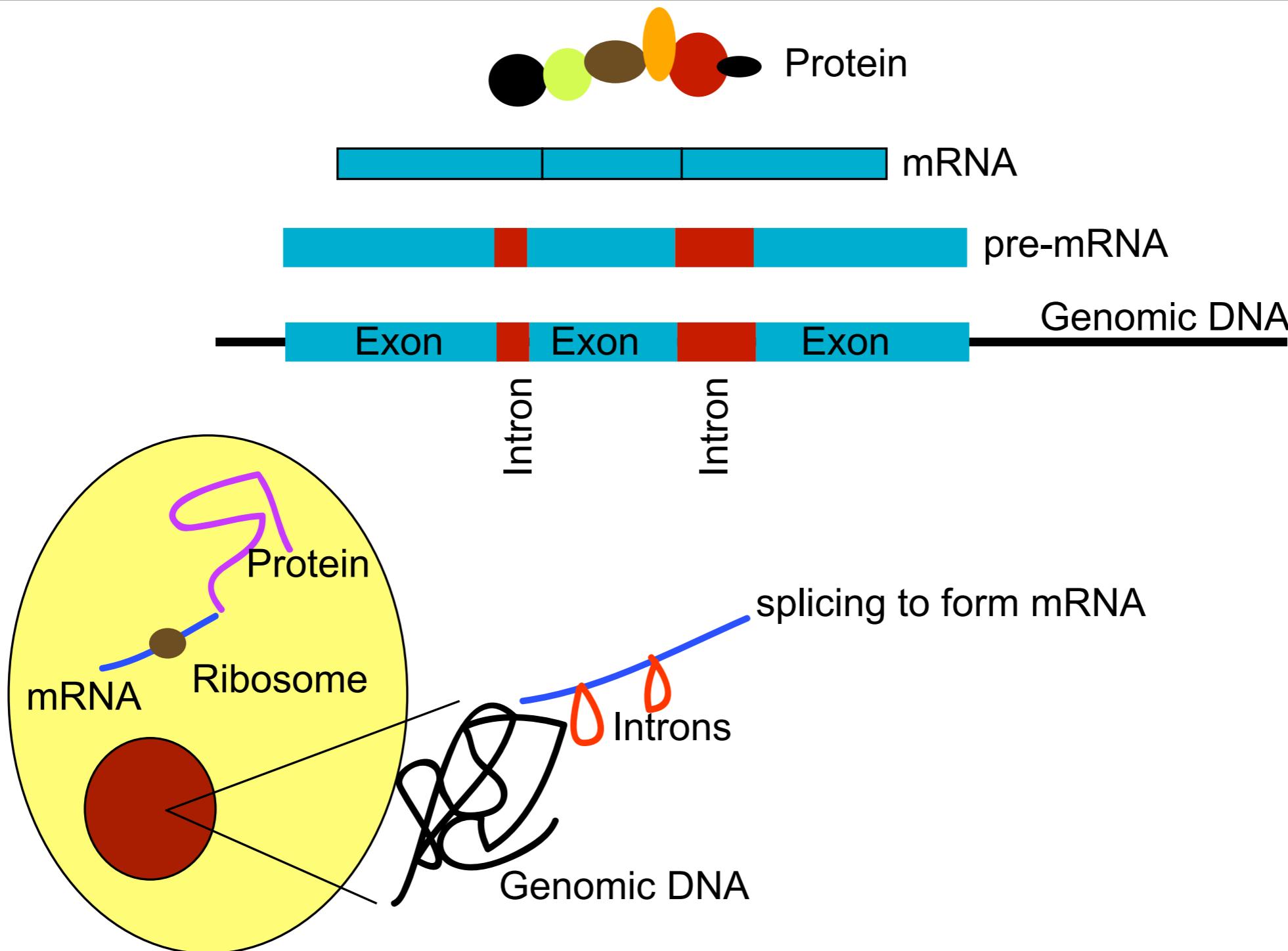
- Smaller than most vertebrate and plant genomes
  - *A. gossypii* 8.5 Mb; *S. cerevisiae* 12 Mb
  - *N. crassa* 40 Mb
  - Animals: 100 Mb worm; 3000 Mb Human
- Vary in protein coding gene content
  - 4,700 in *A. gossypii*; 5,800 in *S. cerevisiae*
  - 16,000 in *R. oryzae* or *S. nodorum*
  - 19,000 in Fruitfly; 25,000 in worm

# Fungal comparative genomics

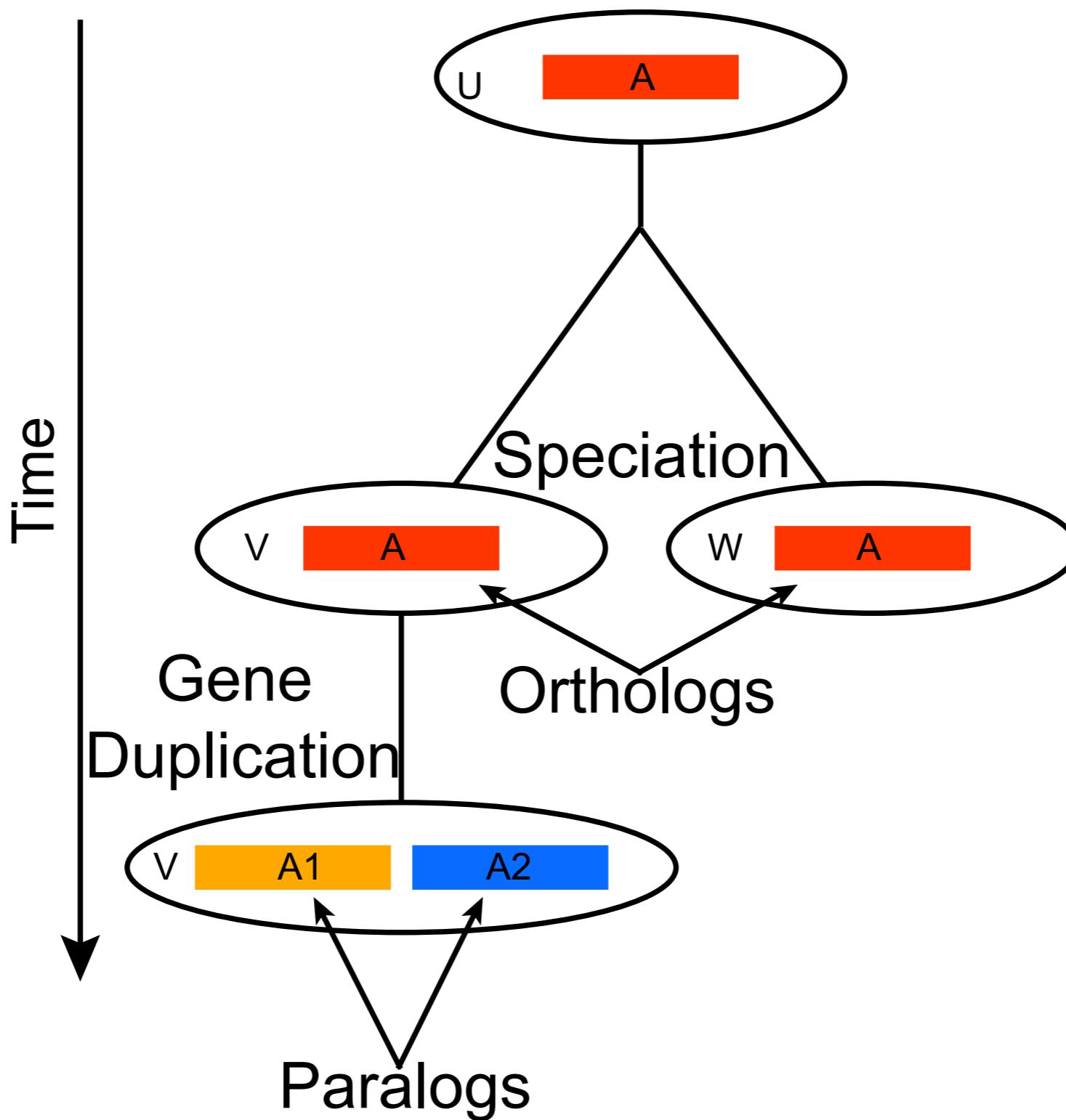


# Central dogma of eukaryotic biology

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# Orthologs and Paralogs

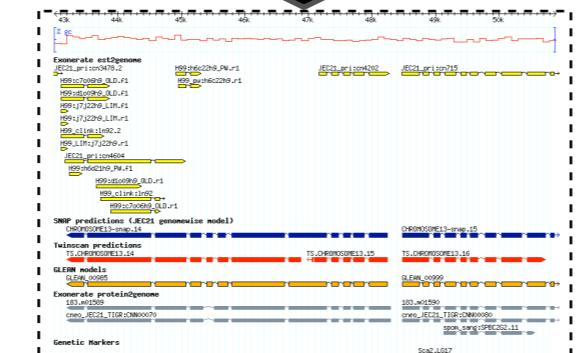
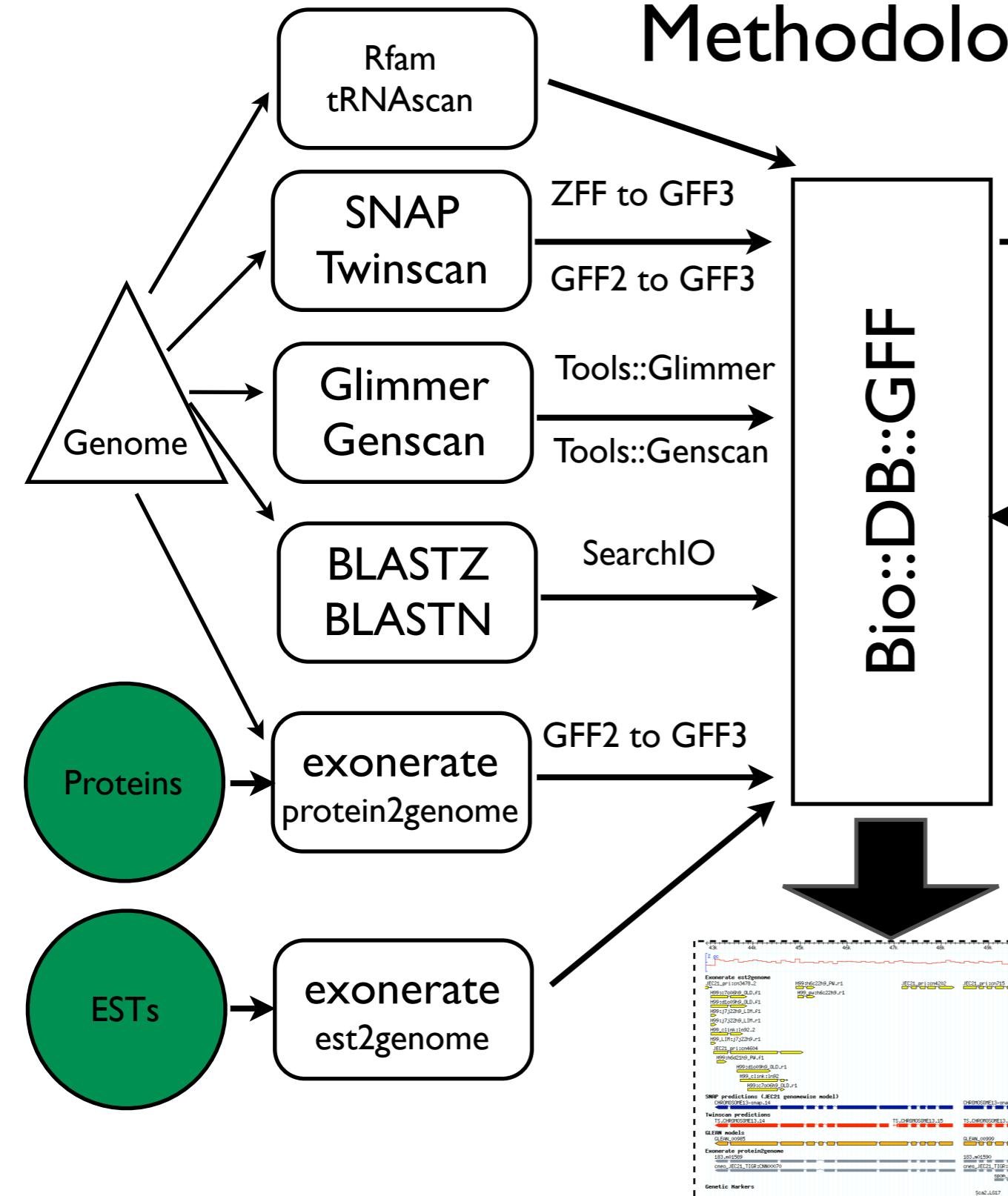


# Genome annotation

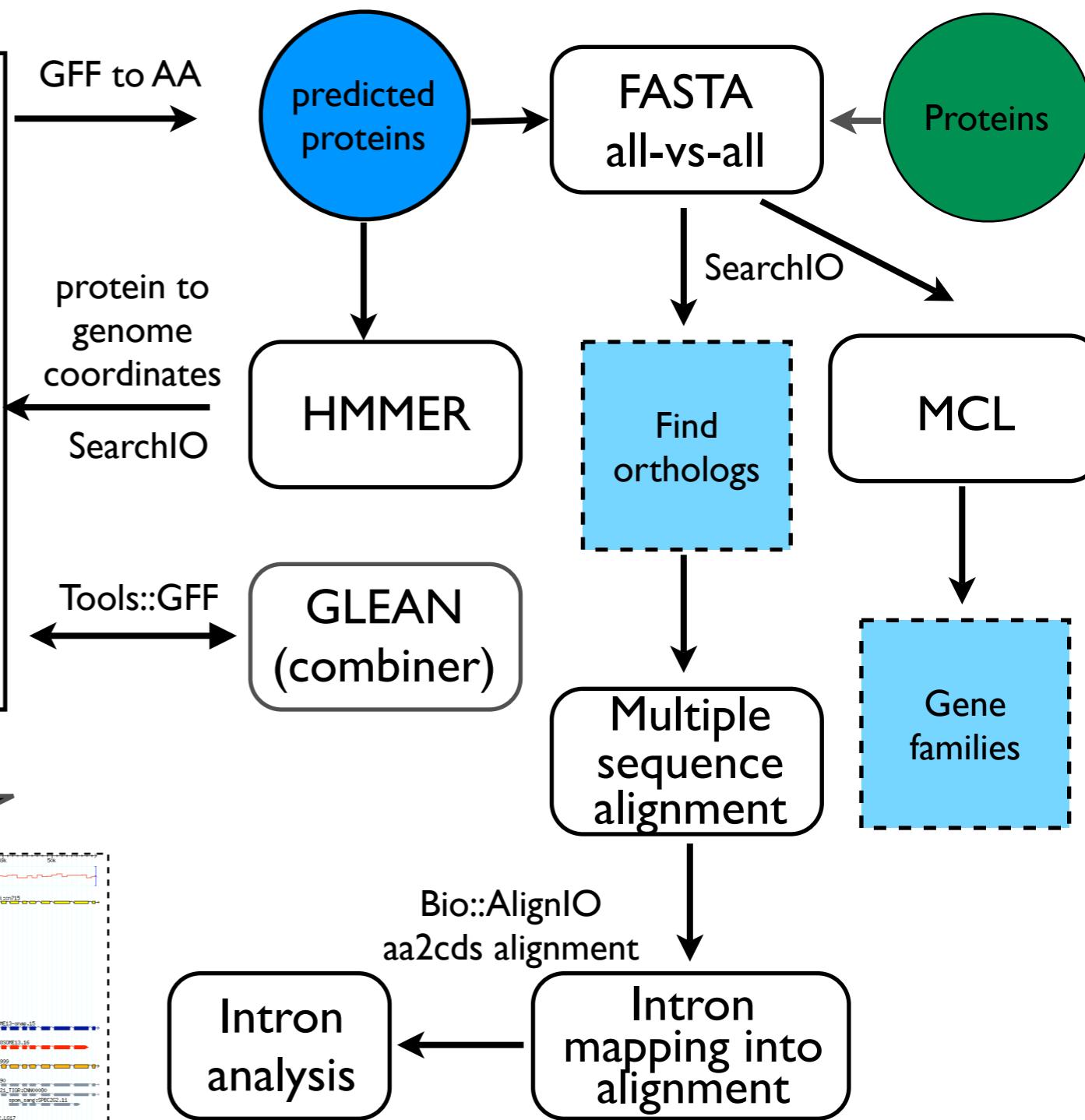
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- Available fungal genomes were only assembled genomic sequence.
- Need systematic and consistent gene predictions for genome comparisons
- Automated annotation pipeline for gene prediction.

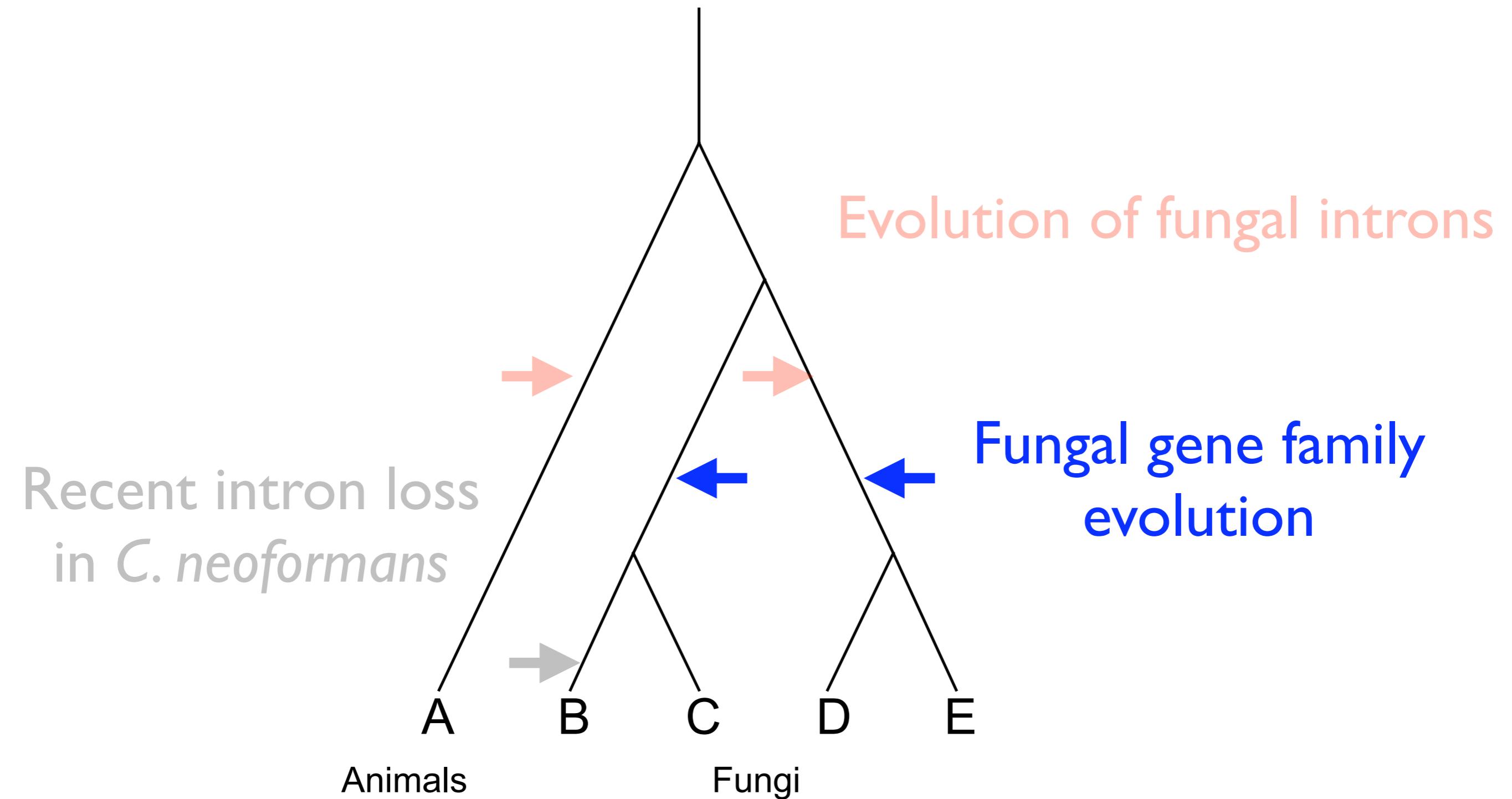
# Methodology



# Analysis



# Fungal comparative genomics



# Gene family evolution

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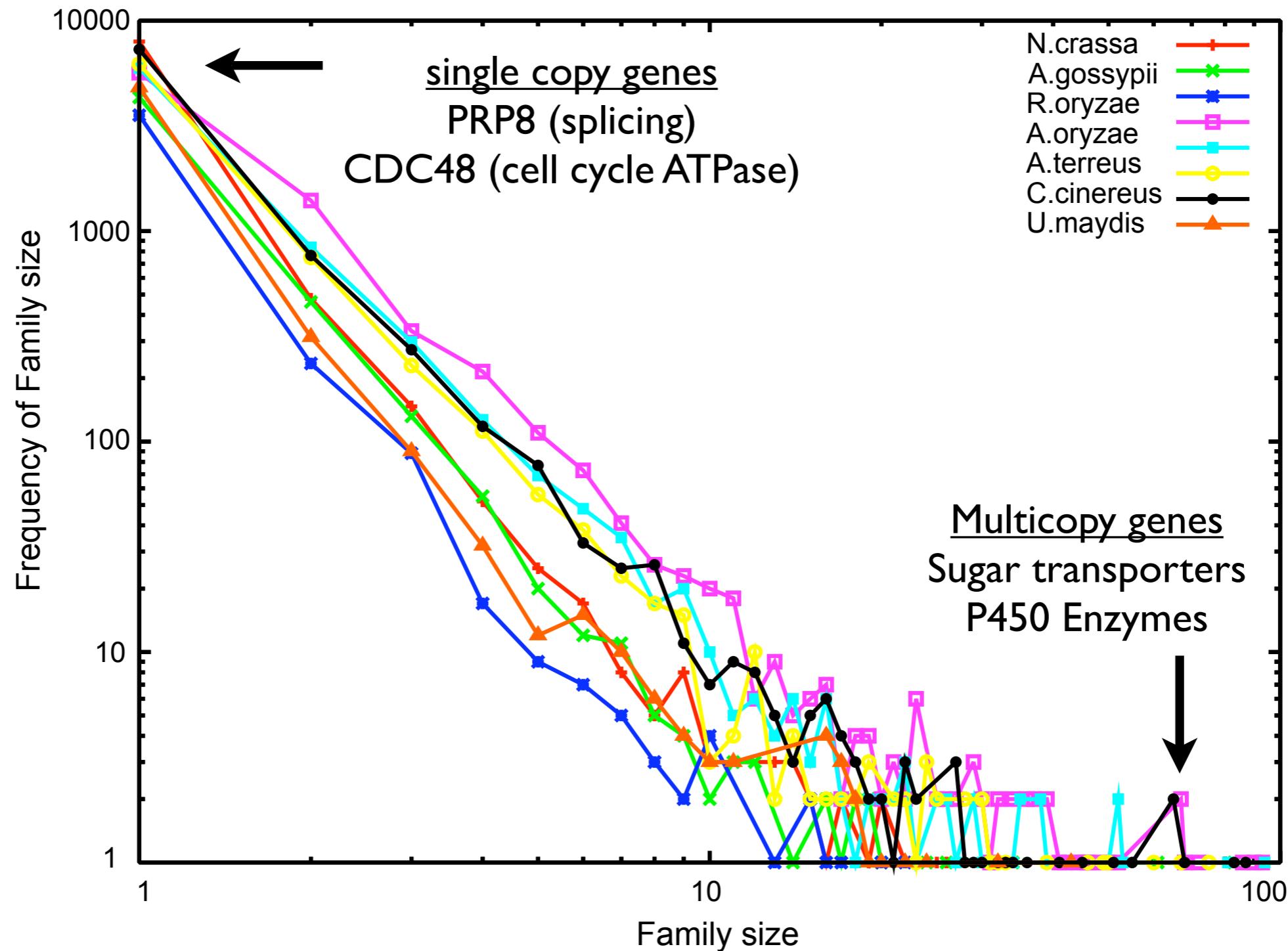
- Gene families are the crucible of new genes and thus new functions
- Signature of adaptive evolution often confounded in multi-gene families
- Can we identify families that have unexpectedly large changes in size across a phylogeny?
  - Follow up these families with more focused studies

# Identifying gene family expansions

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- Previous work only considered pairwise
- Ad hoc comparison of gene family sizes
  - *C.elegans-C.briggsae* - GPCR family expansions (Stein et al, *PLOS Biology* 2004)
  - *A.gambiae-D. melanogaster* - Mosquito specific family expansions related to symbiotic bacteria (Holt et el, *Science* 2002).

# Gene family sizes follow power law distribution

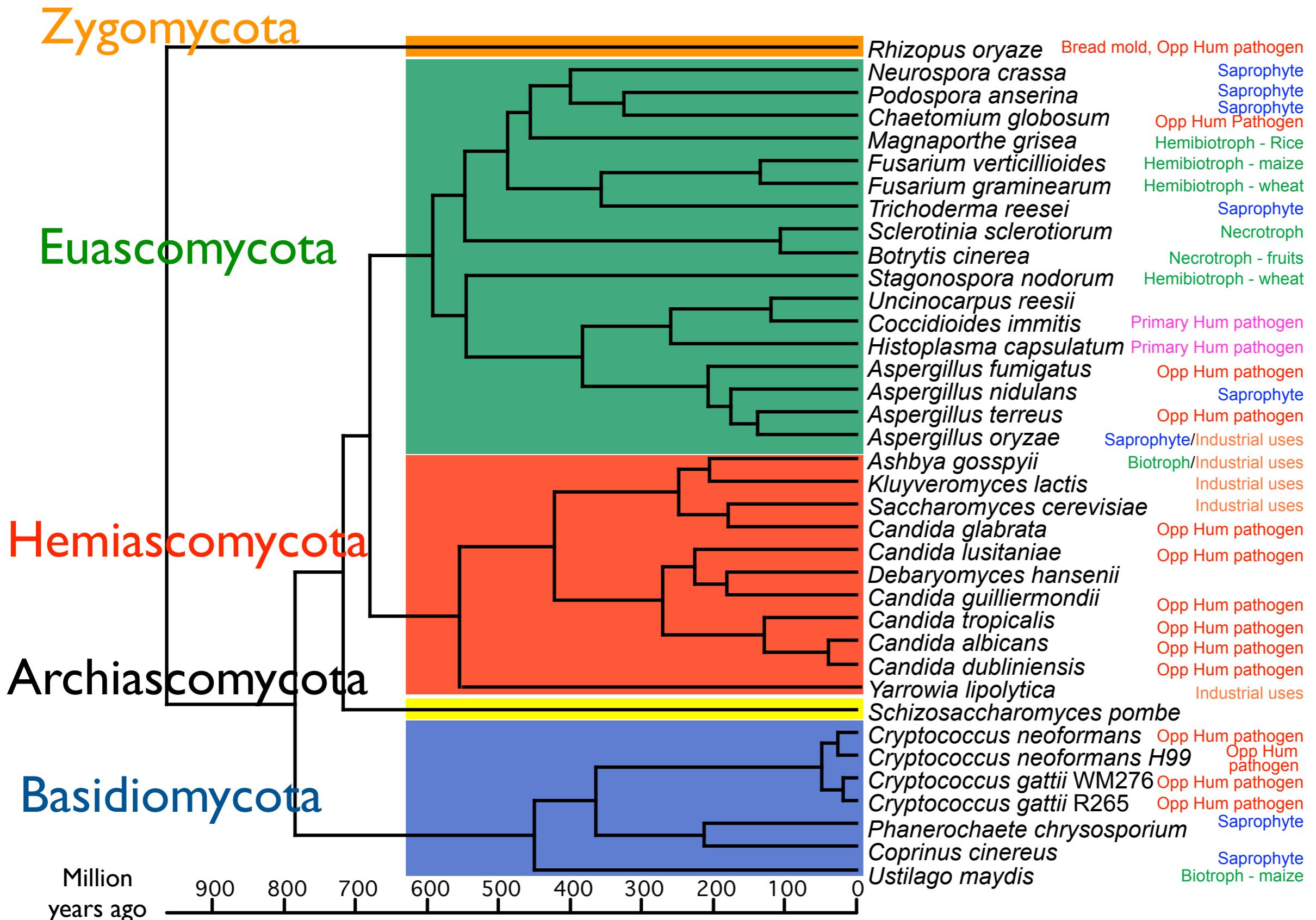


# Phylogenetic evaluation of gene family size change

- Previous methods only used *ad hoc* statistics
- Explicit model for gene family size change according to a Birth-Death (BD) models
- Apply BD to family size along phylogeny using probabilistic graph models
- CAFE - Computational Analysis of gene Family Evolution

Hahn et al, *Genome Res* 2005  
De Bie, et al *Bioinformatics* 2006  
Demuth et al, *submitted*

# Fully sequenced fungal genomes



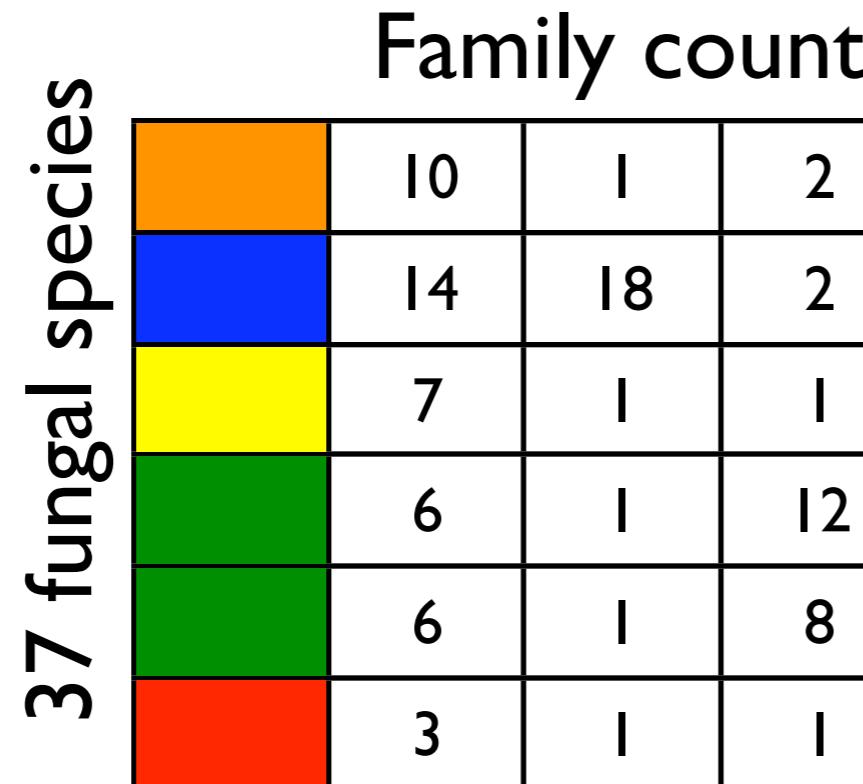
FASTA  
all-vs-all



MCL

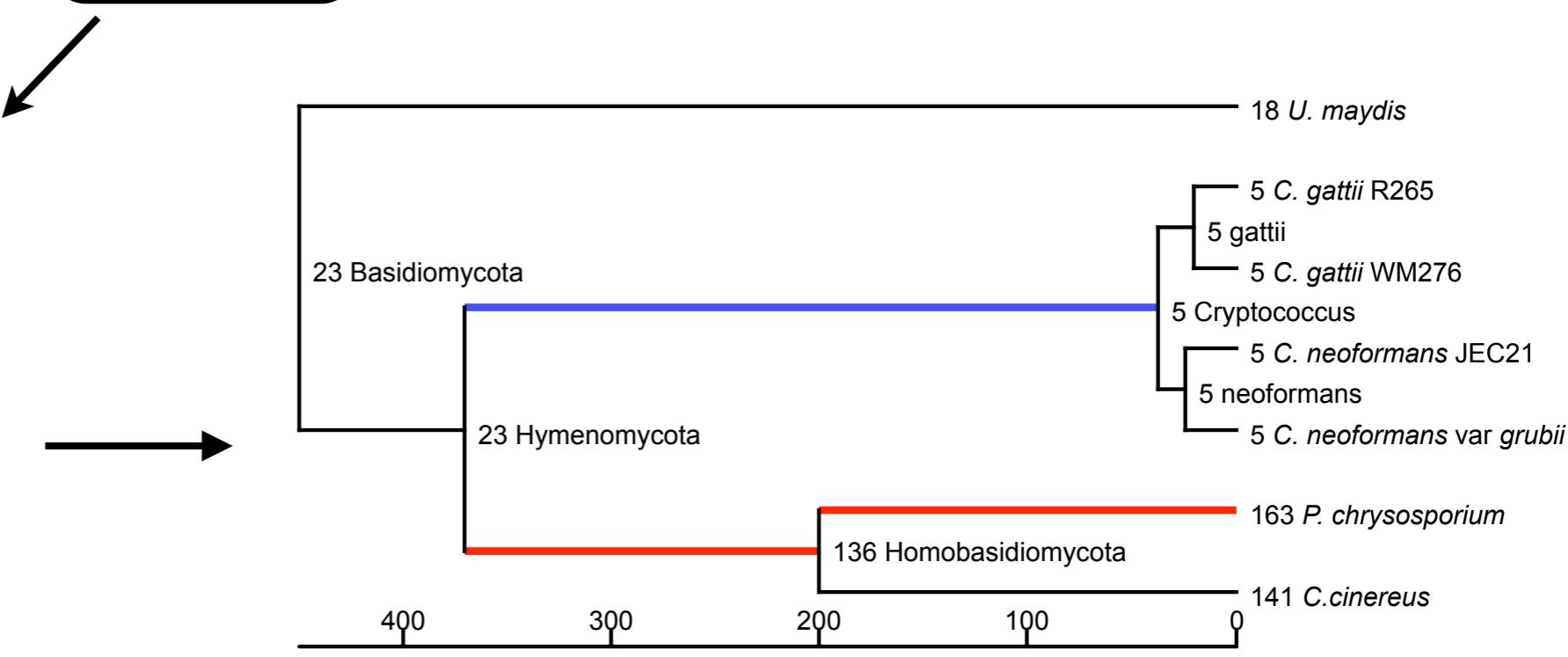


Gene  
families



CAFE

Family 1	P < 0.001	Branch A
Family 2	P < 0.001	Branch B
Family 3	P=0.02	Branch C,E
Family 4	P=0.03	Branch D



# Families with significant expansions

Transporters  
Kinases  
P450  
Oxidation

Vitamin & Cofactor transport	Methytransferase
Lactose & sugar transport	Cytochrome P450: CYP64
Amine transport	Cytochrome P450: CYP53,57A
Myo-inositol, quinate, and glucose transport	Cytochrome P450
Oligopeptide transport	Kinase
ABC transporter	Subtilase family
MFS, drug pump, & sugar transport	NADH flavin oxidoreductase
Transport	Aldehyde dehydrogenase
Monocarboxylate & sugar transport	Aldo/keto reductase
ABC transport	Multicopper oxidase
Amino acid permease	AMP-binding enzyme

# Transporters

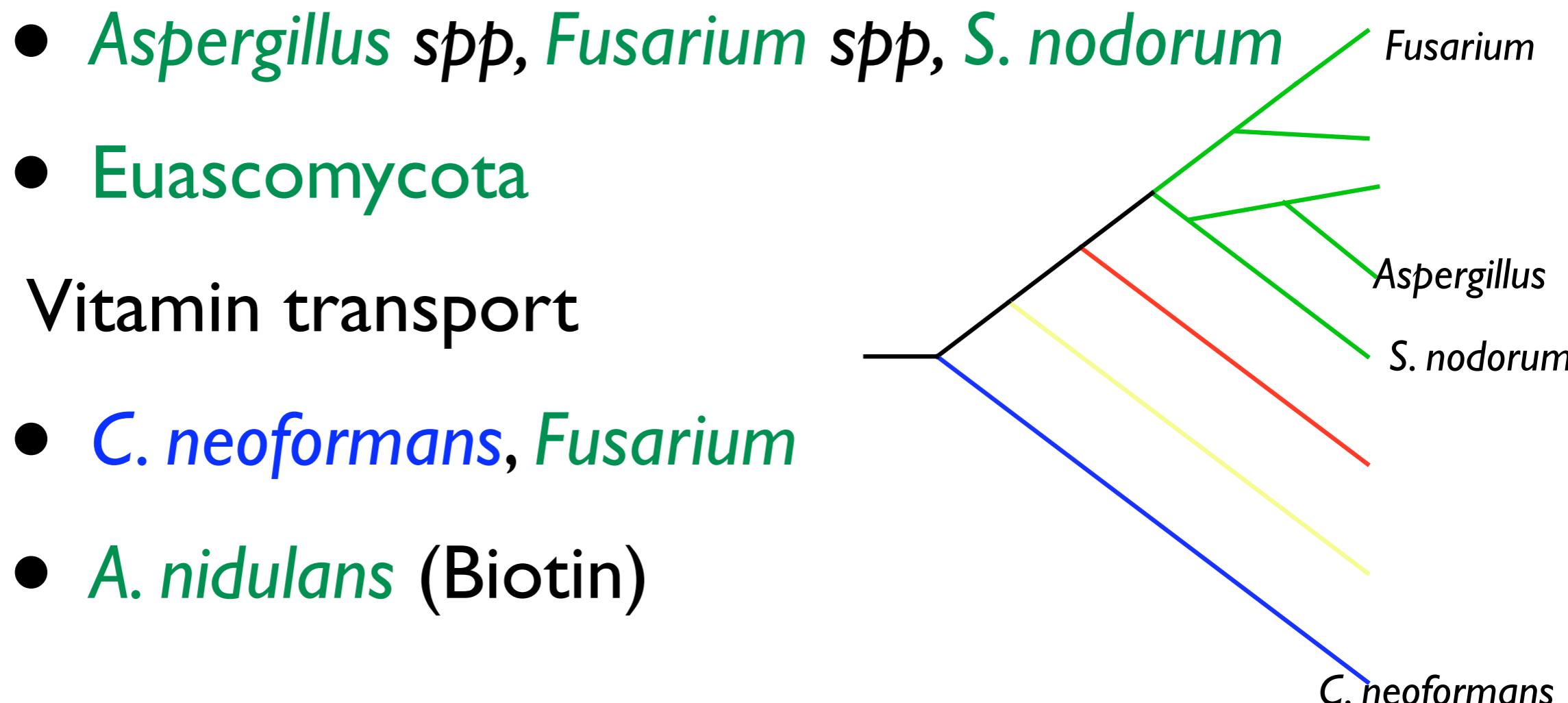
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- Of 45 significant families, 22 were related to transport
- Vitamin and amino acid transport
- Sugar and sugar-like transporters
- Multidrug and efflux pumps
- ABC transporters (ATP Binding Cassette)

# Branches with transporter expansions

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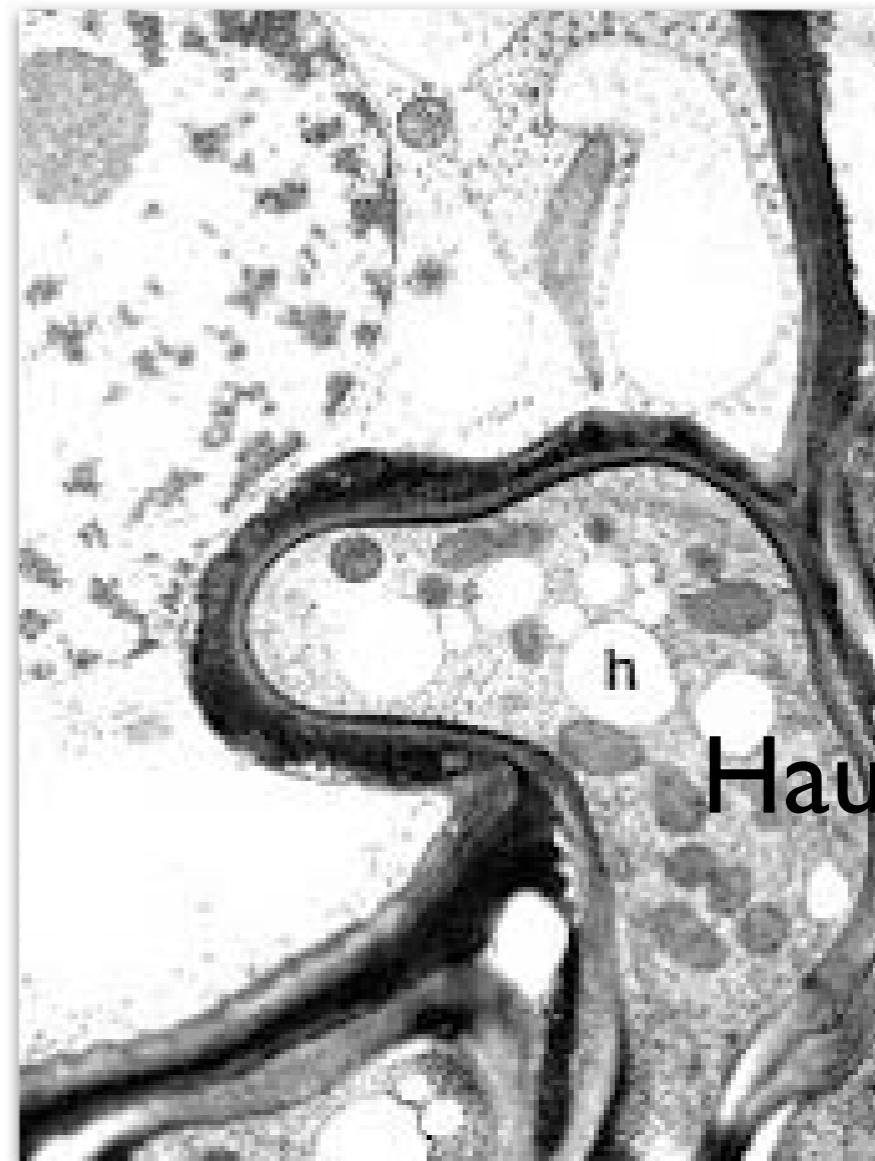
- Sugar related, Drug pump, and Major Facilitator Superfamily



# Sugar transporter use in phytopathogens

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- Sugar transporters are used to extract nutrients from host
- Haustorium: specialized structure for plant parasitism
- Many sugar transporters highly and specifically expressed in haustoria



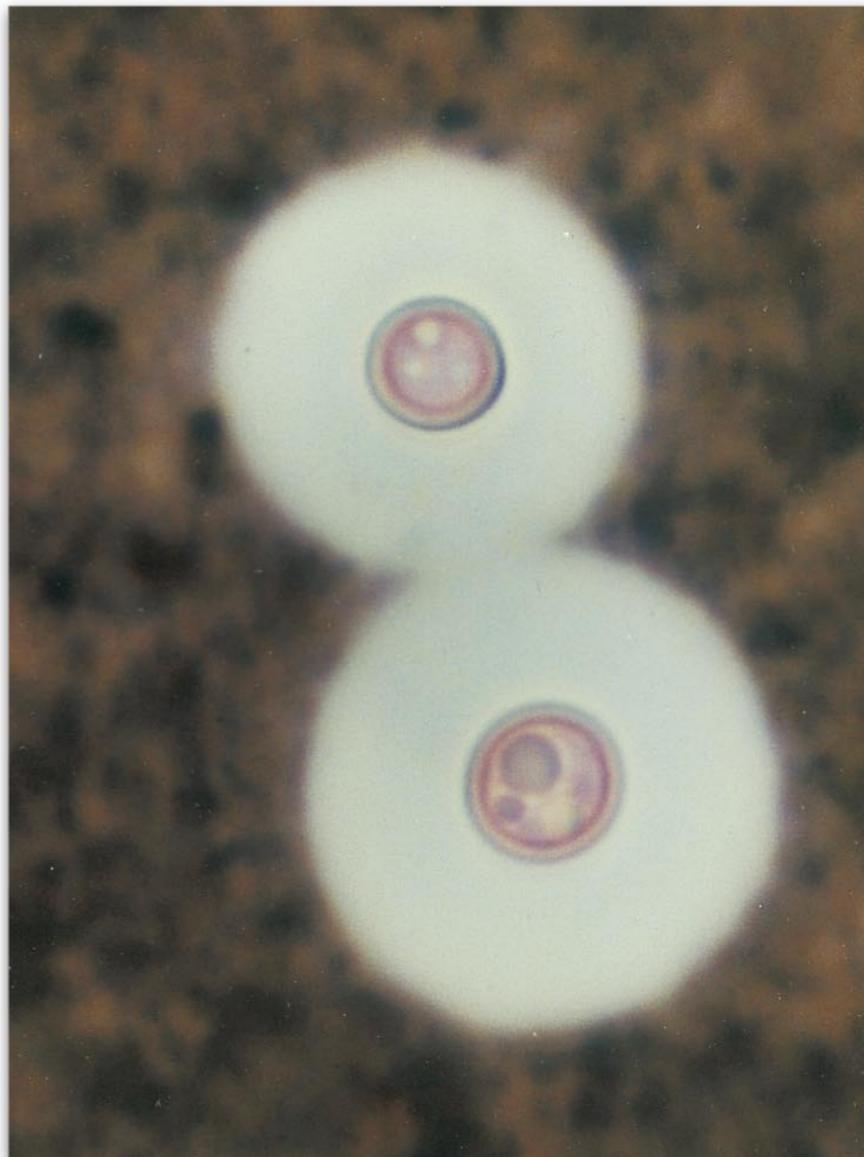
Haustorium

Robert Bauer <http://tolweb.org/>

# *Cryptococcus* sugar transporters

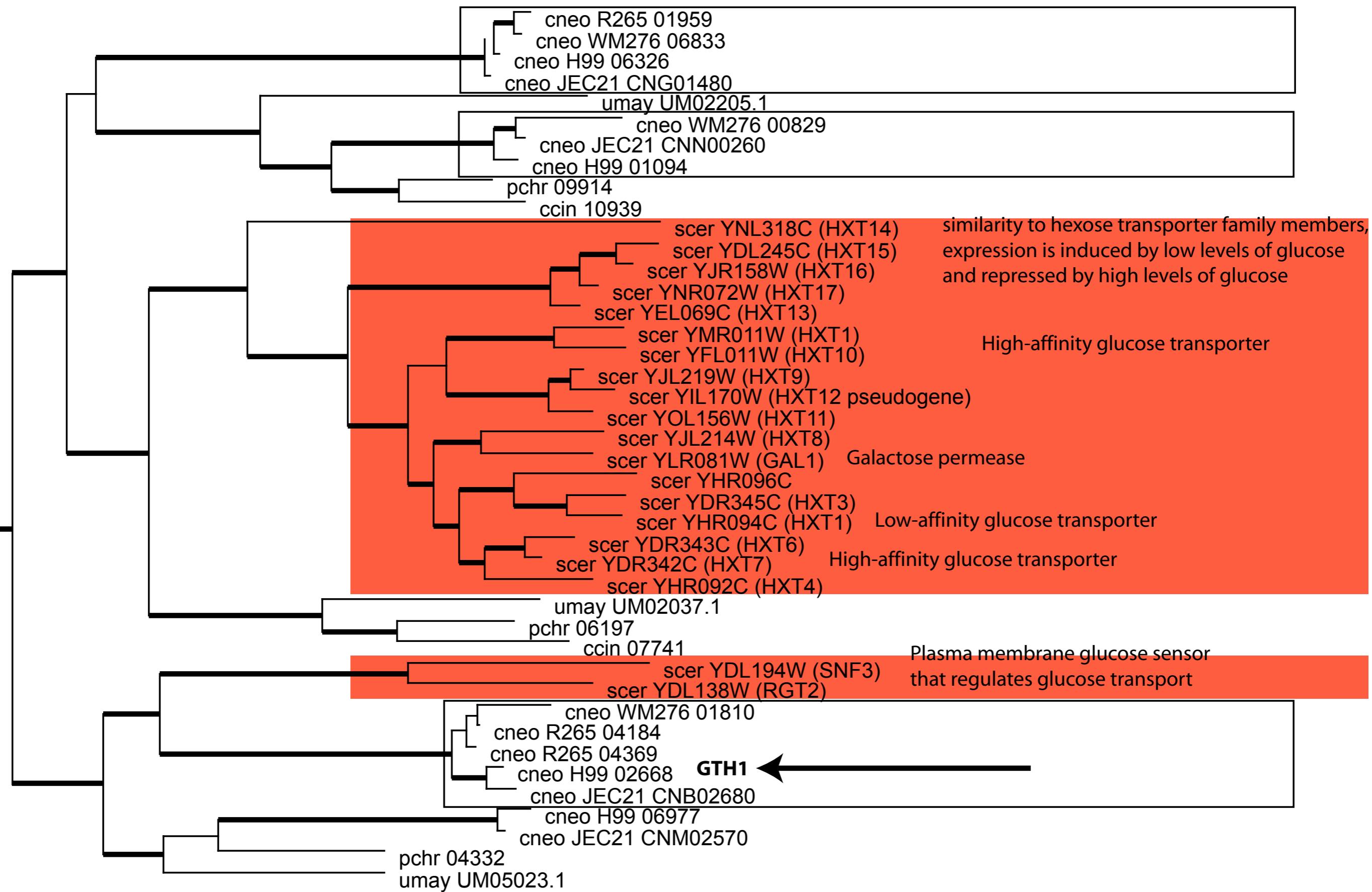
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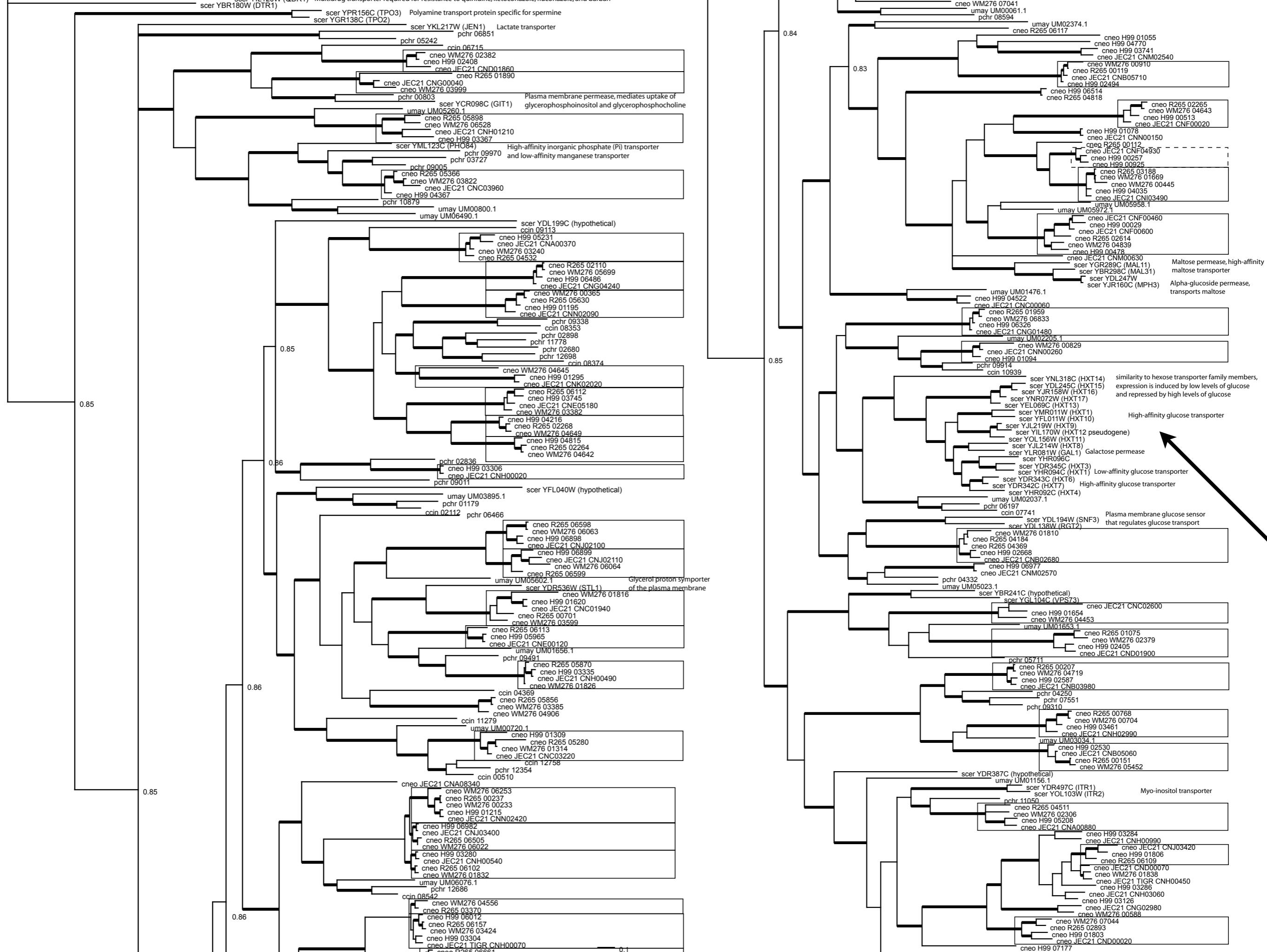
- 3x as many sugar transporters in *C. neoformans* (~50) than other basidiomycetes
- “sugar coated killer”
- Capsule is a mixture of glucose, xylose, and mannose.
- Transporters could be important in capsule synthesis



Zerpa et al, 1996

# **Analysis of sugar transporter sub-family**





# GTH I is a Glucose Sensor

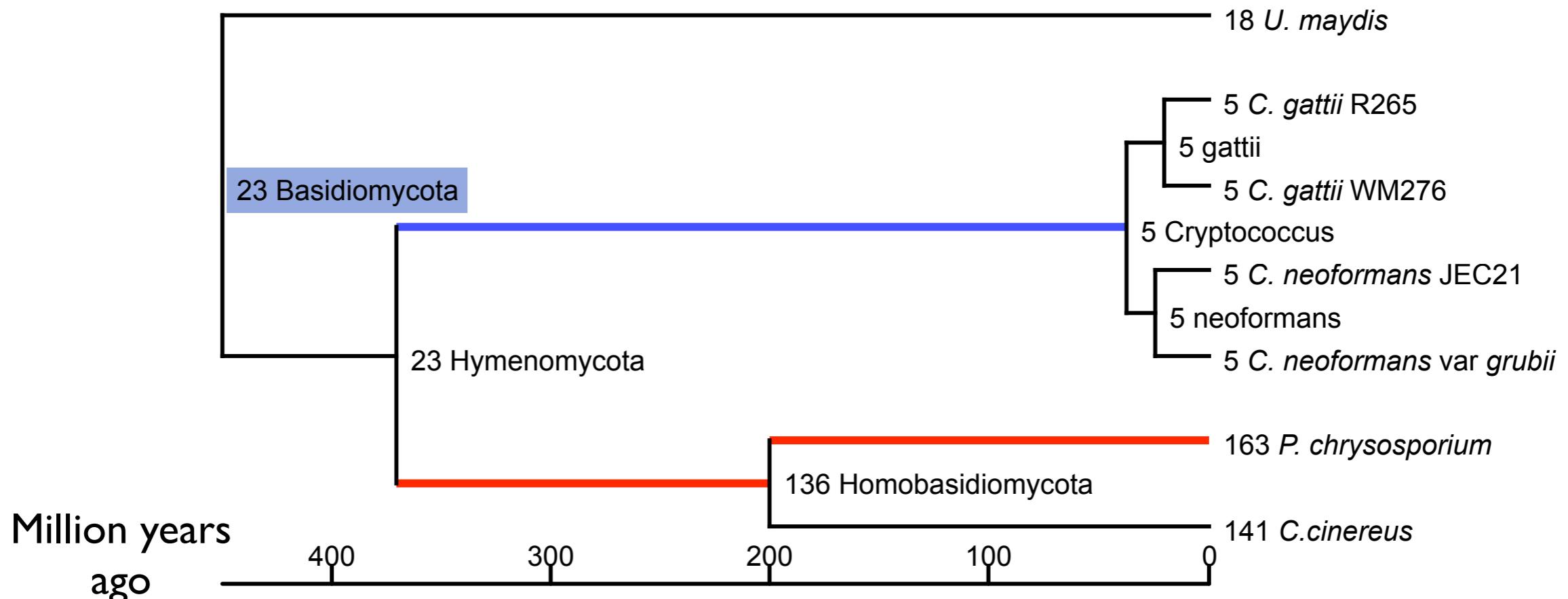
- Melanin production pathway is starvation induced
- GTHI KO is hypermelanized (always glucose starved)
- GTHI overexpression is hypomelanized

Alspaugh et al 1999

# P450 CYP64

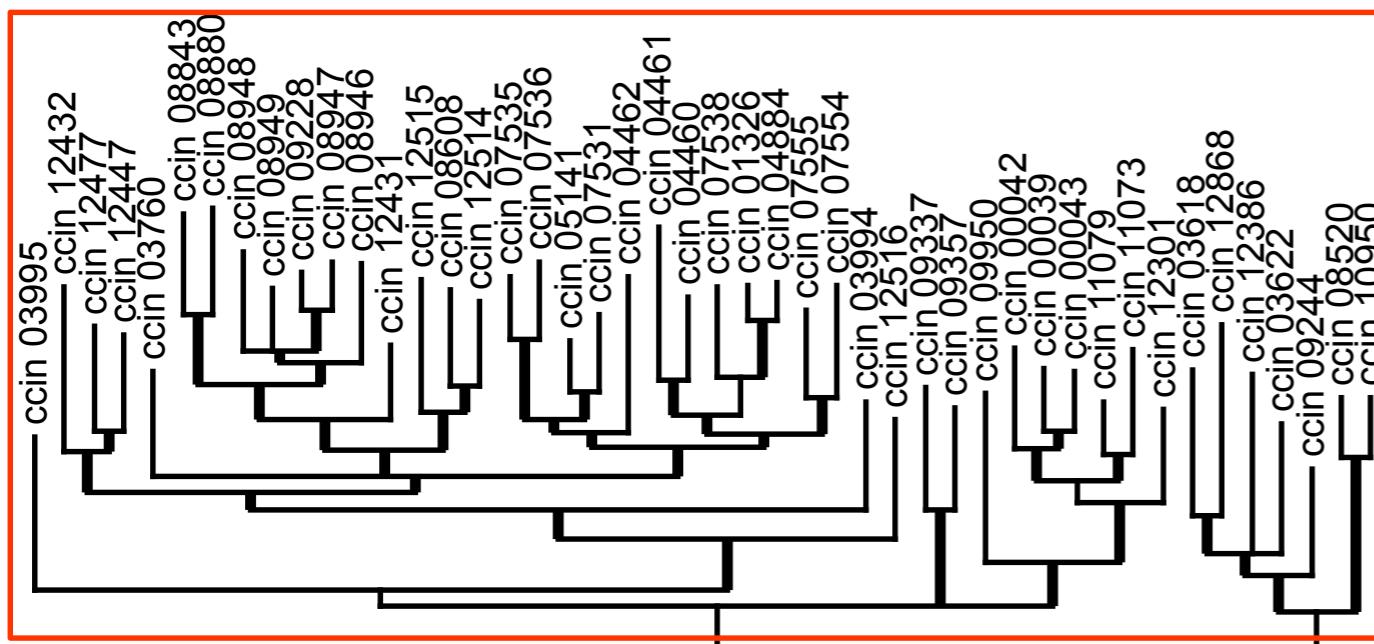
P450 enzymes involved in synthesis and cleavage of chemical bonds. Drug metabolism in animals.

CYP64: Step in *Aspergillus* spp aflatoxin pathway  
*P. chrysosporium* implicated in lignin and hydrocarbon degradation.



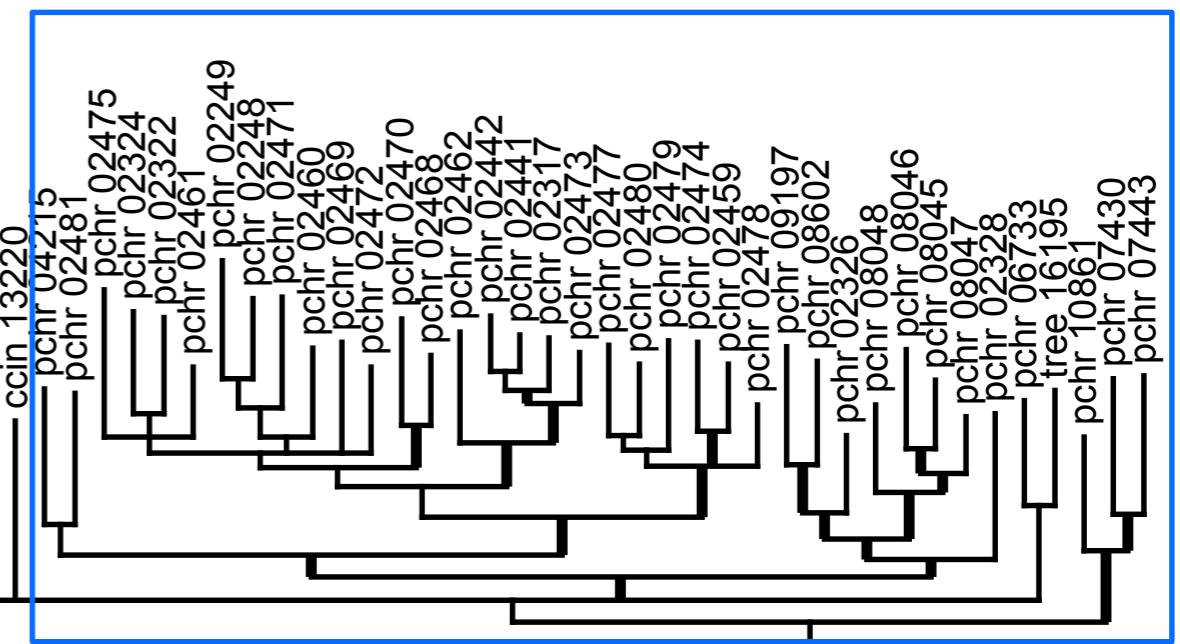
CYP64 was from  
independent duplication

# *C. cinereus* expansion



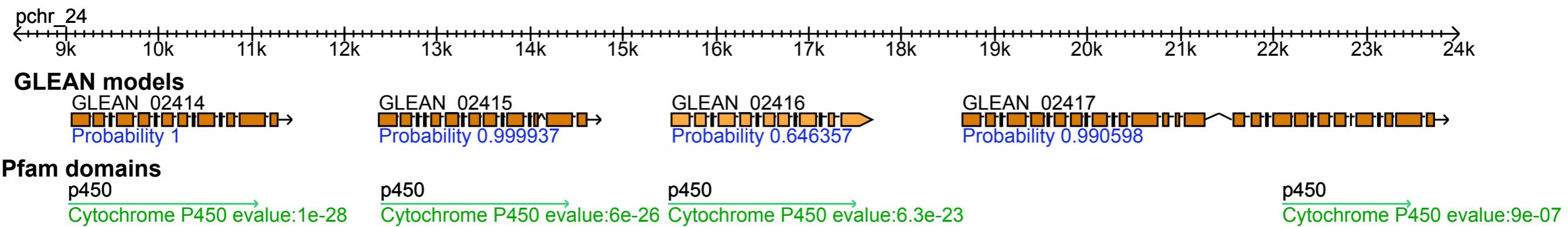
Mario Cervini

# *P. chrysosporium* expansion



Tom Volk

# Local duplications created CYP64 expansion



# Family size contractions

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- *Histoplasma, Coccidioides* many families
- **Hemiascomycetes** - P450
- *C. neoformans* - P450
- *U. maydis* - Lactose transport

# Basidiomycetes

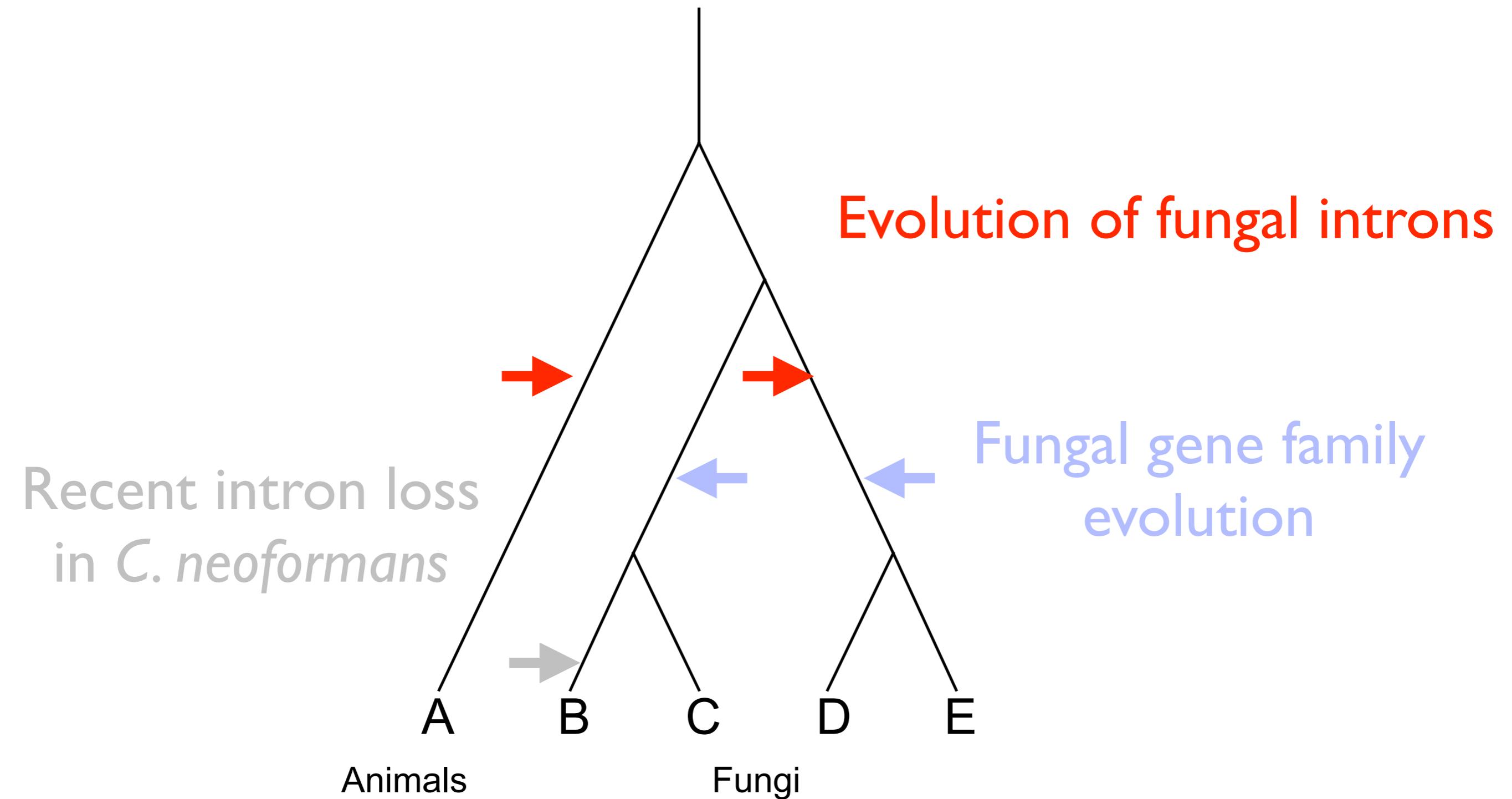
fam	<b>U.maydis</b>	<b>C.cinereus</b>	<b>P.chrysosporium</b>	<b>H99</b>	<b>JEC21</b>	<b>R265</b>	<b>WM276</b>	<b>Cryptococcus &amp; GO Notes</b>	<b>Protein domains</b>
0	1	420	68	140	4	14	70	transposon	Integrase core domain(57);
1	74	108	119	78	81	84	82	kinases. homobasidio	Protein kinase domain(306);
2	44	67	87	48	50	49	52	G-protein coupled receptor protein	WD domain, G-beta repeat(193)
3	42	113	53	27	4	23	106	transposon	Reverse transcriptase
4	18	141	163	5	5	5	5	metal ion binding; cation binding	Cytochrome P450 (39)
5	32	26	76	44	47	46	45	drug transport	Major Facilitator Superfamily(157);
6	17	36	44	47	55	40	39	"hexose transporter	dicarboxylate symporter
7	19	20	23	59	57	47	50	sugar transporter related	Sugar (and other) transporter(167);
8	30	33	64	32	33	31	35	"siderophore biosynthesis?	dihydroxybenzoat
9	35	39	35	33	40	34	36	transmembrane receptor activity;	Ras family(130); ADP-ribosylation
10	32	44	45	29	27	31	30	localization hydrolase	ABC transporter (112); RecF/RecN/
11	5	125	61	11	11	11	13	hydrolyzing O-glycosyl	WSC domain (24); Glycosyl
18	4	2	5	46	43	22	30	oxidoreductase activity;RNA	Oxidoreductase family, NAD-
20	0	109	36	0	0	0	0	kinase activity; phosphotransferas	

# Conclusions

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- Sugar transporters are highly expanded in independent lineages
  - Saprophytic and phytopathogenic lifestyles
- P450 CYP64 independent expansions in **Homobasidiomycetes**
  - Lignin degradation and saprophytic lifestyles
  - Family size contractions among lineages containing primary pathogens
  - Genome streamlining?

# Fungal comparative genomics



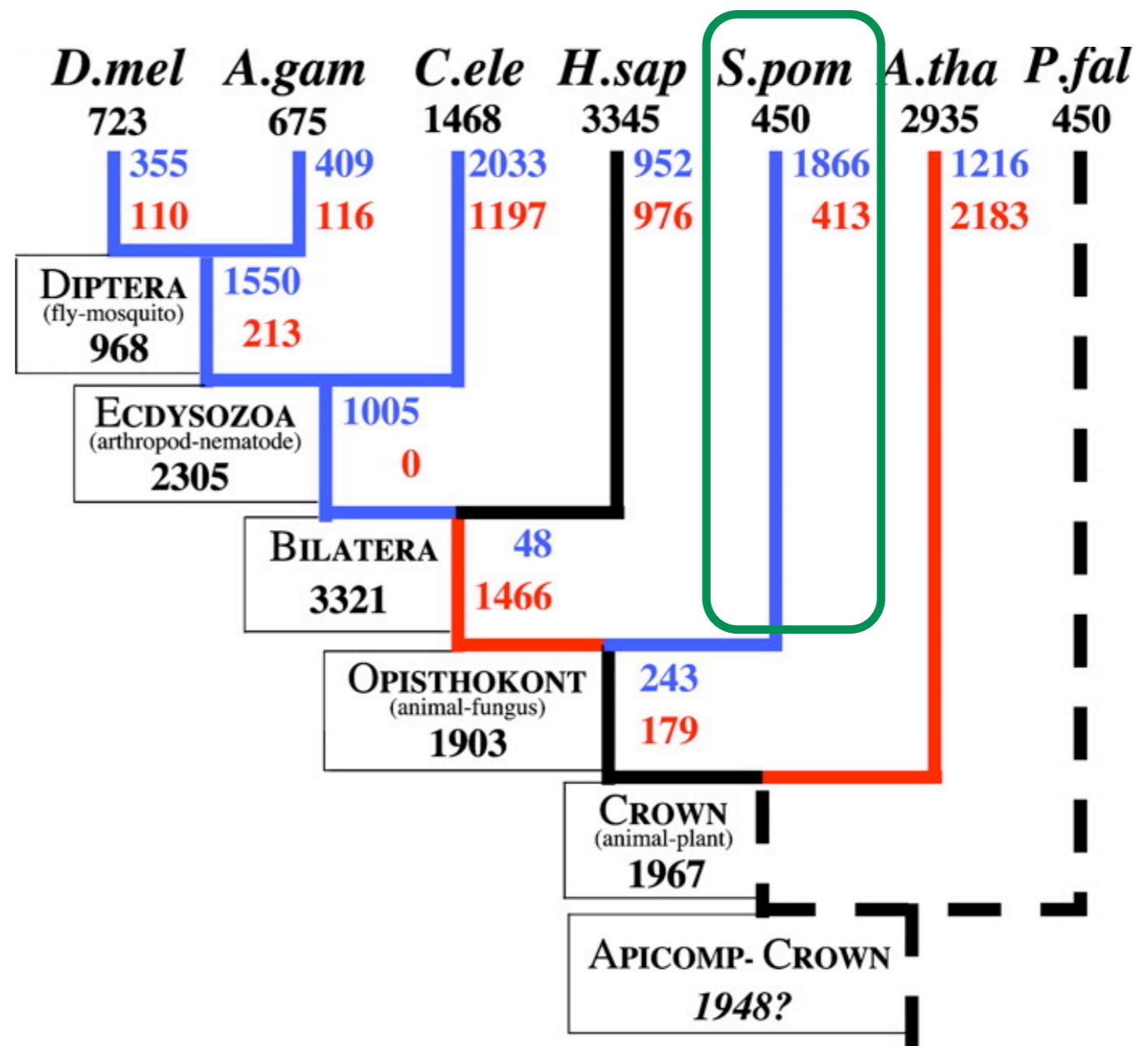
# Evolution of gene structure

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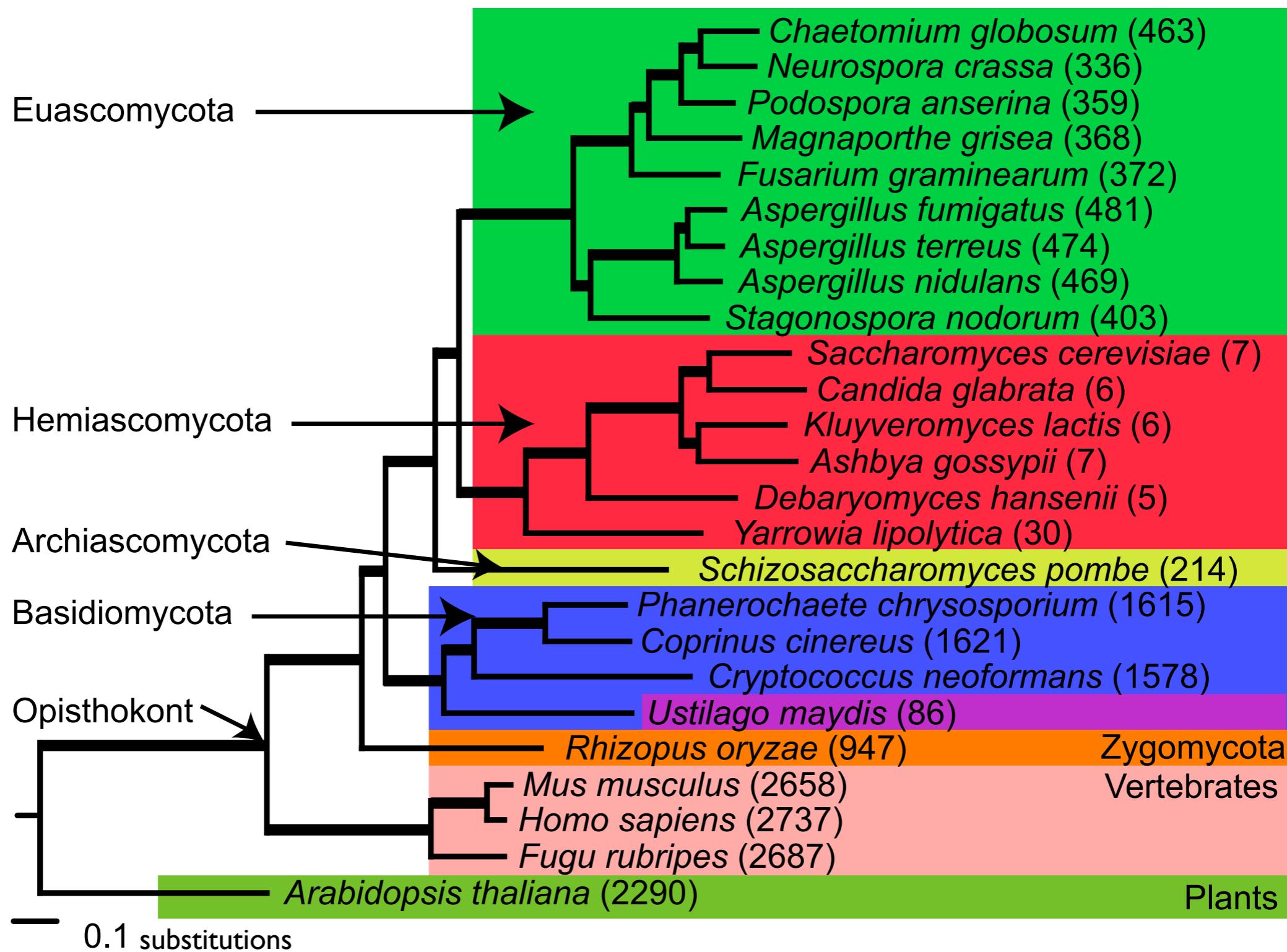
- Present day introns
  - Recent insertions?
    - Introns late hypothesis
  - Present in eukaryotic ancestor?
    - Introns early hypothesis / exon theory of genes
  - Mixture of two?

# Previous work on intron evolution

- Rogozin et al. 2003
  - 7 genomes
  - 684 genes, 7236 positions
  - Parsimony analysis
- Analysis methods
  - Roy and Gilbert. 2005
  - Csűrös. 2005
  - Nguyen et al. 2006

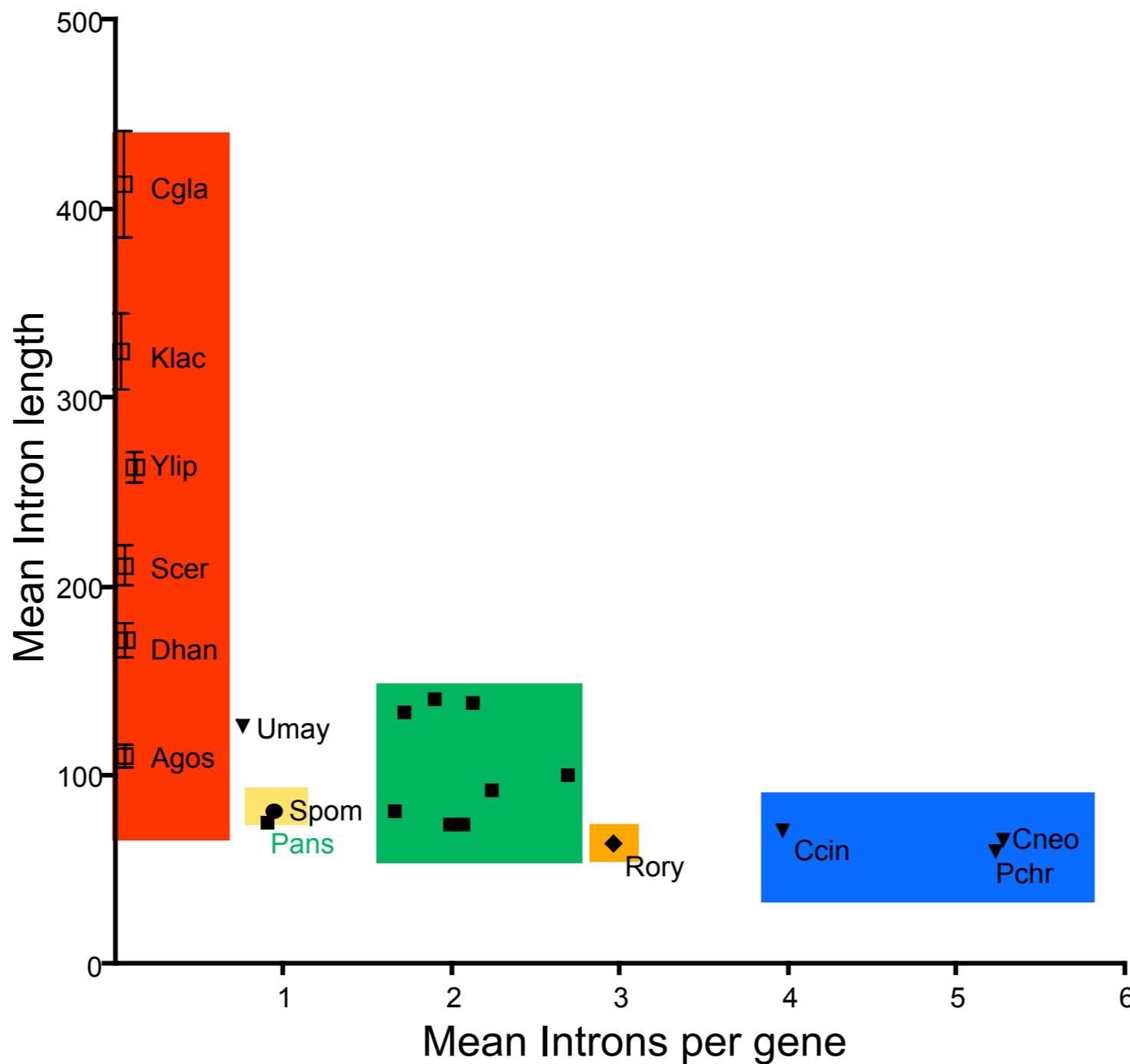


# Calculating intron densities across a phylogeny



# Intron frequency varies among the fungi

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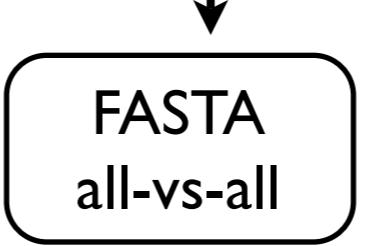
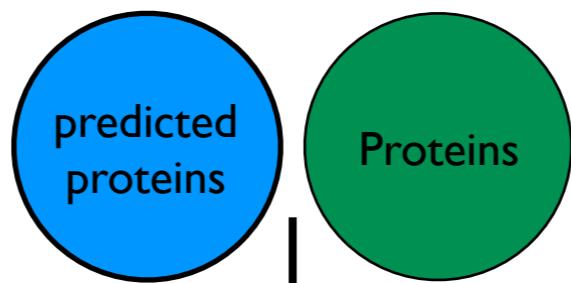
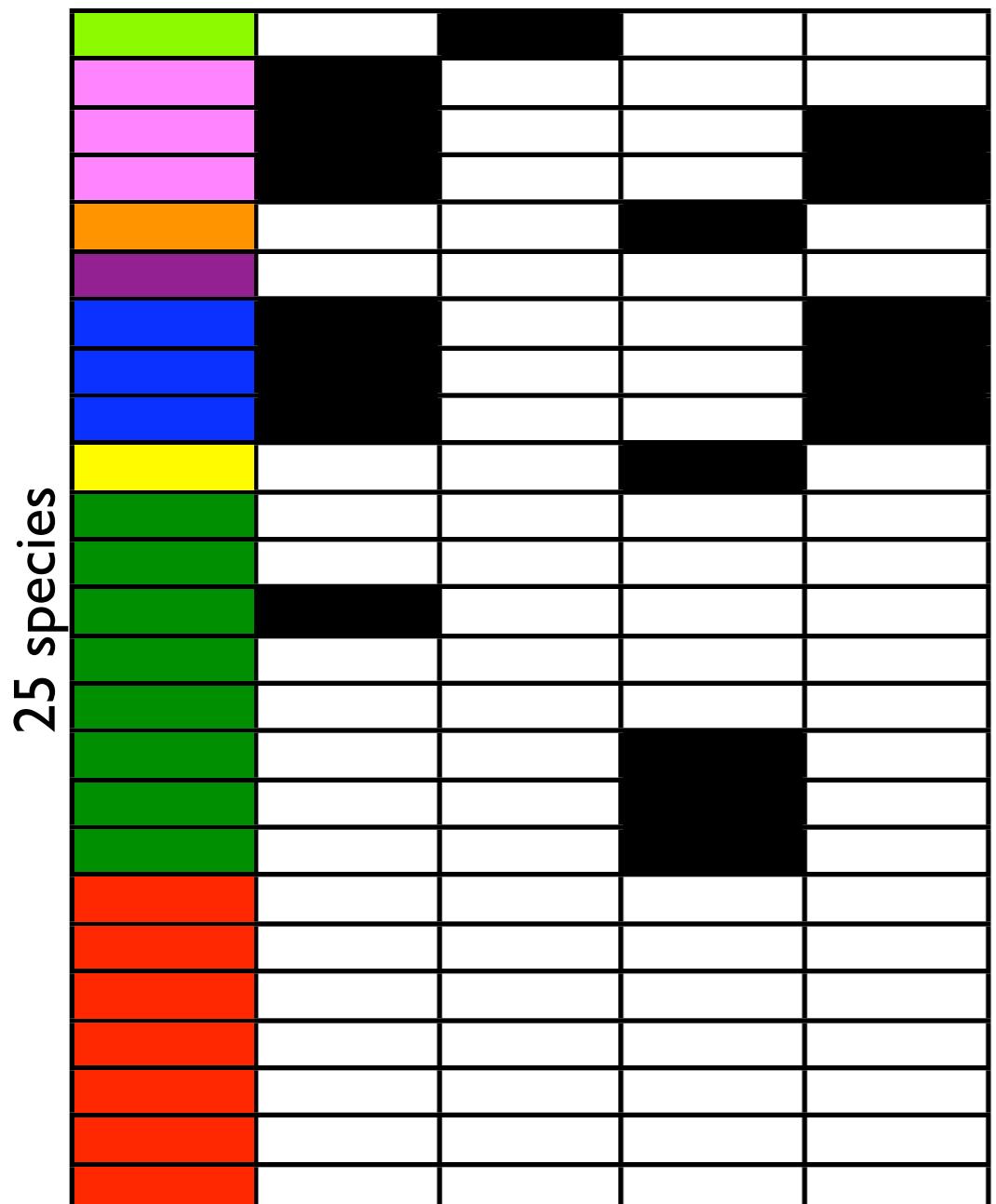
# Analysis of whole genomes

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- 25 entire genomes
  - 21 fungi, 3 vertebrates, 1 plant
- Largest dataset ever assembled for intron analysis
- 1160 orthologous genes
- 7533 intron positions
- 4.15 Mb conserved coding sequence per genome

# Analysis methods

7533 intron positions

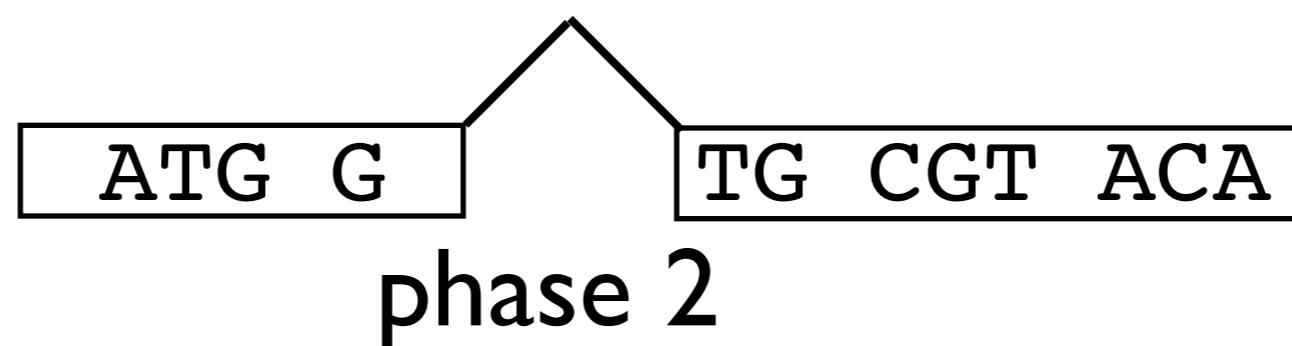
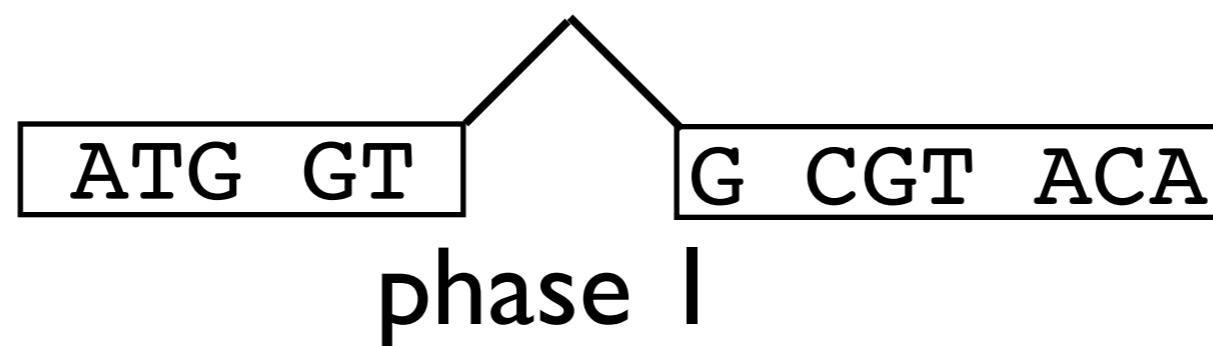
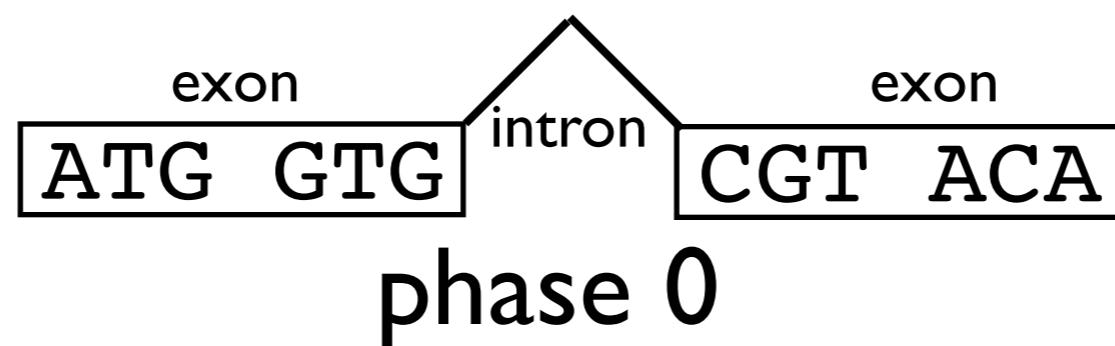


<i>A.thaliana</i>	TWLPEQVA-FIQS1MG-NDKANS-YW
Fugu	QWTQEQQVQ0SVQE-MG-NAKAKR-LY
Human	QWTQEQQI0CMQE-MG-NGKANR-LY
Mouse	QWTQEQQI0CMQE-MG-NGKANR-LY
<i>R.oryzae</i>	TWVQDQVE-NMVR-WG-NERANK2YW
<i>U.maydis</i>	TWTREQVD-RMKE-VG-NLKNR-KY
<i>C.neoformans</i>	IWTPEQME0SIQK-WG-NKRANM-YW
<i>C.cinereus</i>	VWTPEQME0SIQK-WG-NRRANL-YW
<i>P.chrysosporium</i>	VWTVEQMN0SIQK-WG-NKRANI-YW
<i>S.pombe</i>	SWTDEQTE-NMTR-WG-NERANL2YW
<i>F.graminearum</i>	SWTDEQLQ-SVLK-WG-NARANK-YW
<i>N.crassa</i>	AWTDEQLQ-SVLN-WG-NARANK-YW
<i>C.globosum</i>	SWTNEQVD0NMRK-VG-NVVSNK-LY
<i>P.anserina</i>	SWTDEQLQ-SVLN-WG-NARANK-YW
<i>M.grisea</i>	SWTDEQLQ-SVLS-WG-NARANK-YW
<i>S.nodorum</i>	TWTDEQLQ-SVLK-WG-NARANK-YW
<i>A.nidulans</i>	AWTDEQLQ-SVVR-WG-NARANK2YW
<i>A.fumigatus</i>	SWTDEQLQ-SVIK-WG-NARANK2YW
<i>A.terreus</i>	SWTDEQLQ-SVLR-WG-NARANK2YW
<i>Y.lipolytica</i>	AWTEEQLA-SMMK-WG-NTRCNM-FW
<i>D.hansenii</i>	AWTDEQVE-LMIK-WG-NEKCI-YW
<i>K.lactis</i>	TWTEEHLE-AVLE-FGNNKKFNE-YY
<i>A.gossypii</i>	TWKEEHLQ-QVVR-FGNNQQQANK-VF
<i>C.glabrata</i>	TWKEEHLK-ELVQ-MRNNVNANR-VY
<i>S.cerevisiae</i>	TWKEEHLV-KLIQ-FKNNLRANS-YY

\* : : : \* :

# Intron phase

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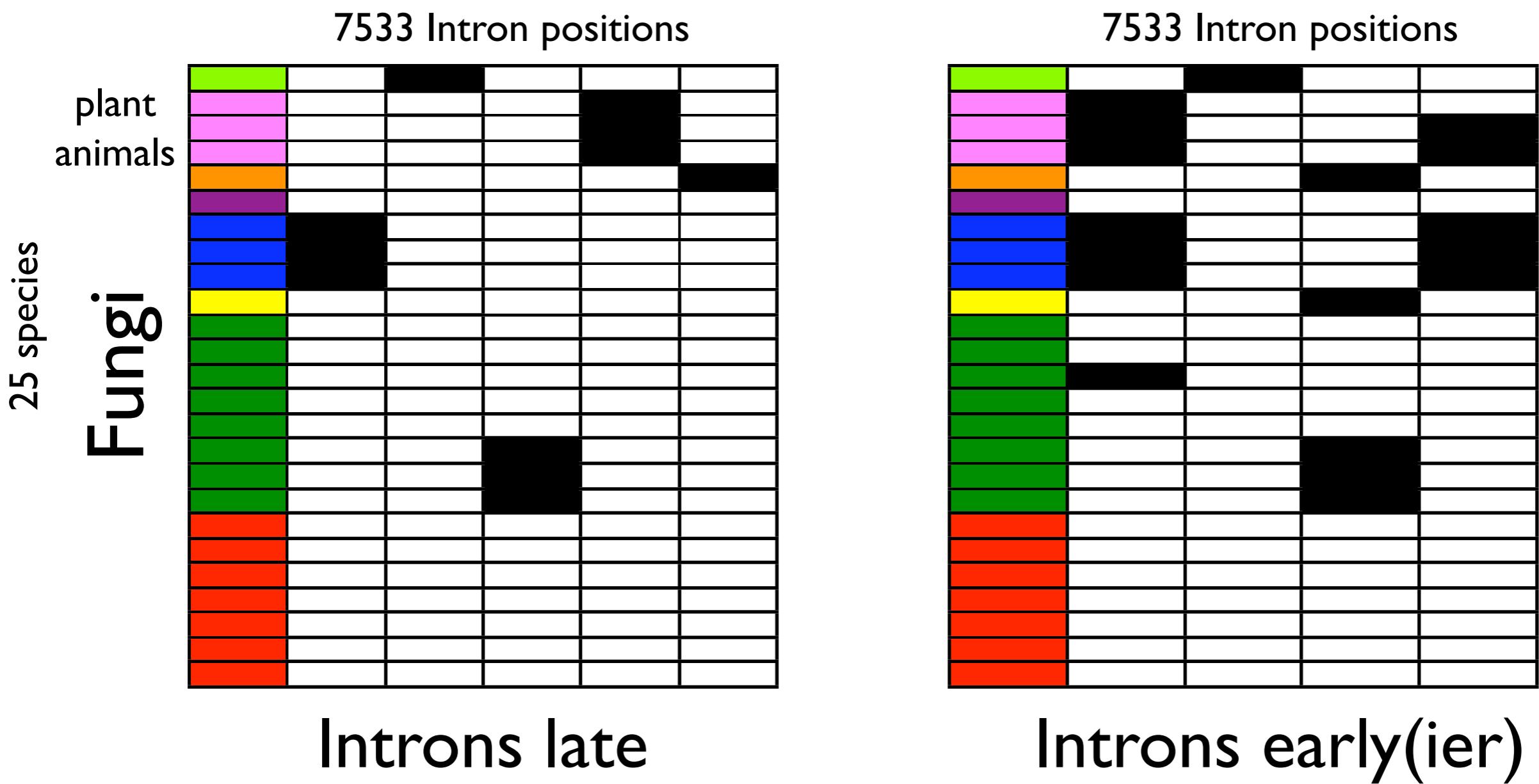


# Conserved intron positions

	TWLPEQVA-FIQS1MG-NDKANS-YWEA-----ELPP-----NYD-----RV-GIENFIRAK2Y-----EEKRWV--
A.thaliana	WTWPEQVA-FIQS1MG-NDKANS-YWEA-----ELPP-----NYD-----RV-GIENFIRAK2Y-----EEKRWV--
Fugu	QWTQEQQVQ0SVQE-MG-NAKAKR-LYEA-----FLPK-----CFQRPETDQ-SAEIFIRDK-Y-----DKKKYMDK-
Human	QWTQEQQI0CMQE-MG-NGKANR-LYEA-----YLPE-----TFRRPQIDP2AVEGFIRDK-Y-----EKKKYMDR-
Mouse	QWTQEQQI0CMQE-MG-NGKANR-LYEA-----YLPE-----TFRRPQIDP2AVEGFIRDK-Y-----EKKKYMDR-
R.oryzae	TWVQDQVE-NMVR-WG-NERANK2YWEA-----NL-----GDRKPS-ES-NMEMWIRAK-Y-----EQKRWA-
U.maydis	TWTREQVD-RMKE-VG-NLKSNR-KYNPDEMNRNPPT-----NMEESERDS-ELEKYIRRK-Y-----EFRRFV-
C.neoformans	IWTPEQME0SIQK-WG-NKRAM-NYER-----HLKA-----GHI-PS-DH2KIESFIRSK-Y-----ETRRWA-
C.cinereus	VWTPEQME0SIQK-WG-NRRANL-YWEA-----HLKP-----GHN-PP-EH2KMESFVRSK-Y-----ESRRWA-
P.chrysosporium	VWTVEQM0SIQK-WG-NKRANI-YWEA-----HLKA-----GHI-PP-DH2KMESFIRSK-Y-----ESKRWA-
S.pombe	SWTDEQTE-NMTR-WG-NERANL2YWEA-----KLAG-----GHV-PS-DS2KIATFIKTK-Y-----EFKKWV-
F.graminearum	SWTDEQLQ-SVLK-WG-NARANK-YWEA-----KLAA-----GHA-PS-EA-KIENFIRTK-Y-----ELKRWV-
N.crassa	AWTDEQLQ-SVLN-WG-NARANK-YWEA-----KLAQ-----GHV-PS-ES-KIENFIRTK-Y-----ELKRWV-
C.globosum	SWTNEQVD0NMRK-VG-NVVSNK-LYNPDN---KNPPVPIDADEA---DG-AMERFIRQK-YIARTLSIGKRRPGGD-
P.anserina	SWTDEQLQ-SVLN-WG-NARANK-YWEA-----KLAP-----GHV-PS-EA-KIENFIRTK-Y-----ELKRWV-
M.grisea	SWTDEQLQ-SVLS-WG-NARANK-YWES-----KLAA-----GHA-PS-EA-KIENFIRTK-Y-----ELKRWV-
S.nodorum	TWTDEQLQ-SVLK-WG-NARANK-YWEA-----KLAP-----GHV-PS-EA-KIENFIRTK-Y-----ESKRWT-
A.nidulans	AWTDEQLQ-SVVR-WG-NARANK2YWEA-----KLAP-----GHV-PP-EA2KIENFIRTK-Y-----ESKRWV-
A.fumigatus	SWTDEQLQ-SVIK-WG-NARANK2YWEA-----KLAP-----GHV-PS-EA2KIENFIRTK-Y-----ESKRWV-
A.terreus	SWTDEQLQ-SVLR-WG-NARANK2YWEA-----KLAP-----GHV-PS-EA2KIENFIRTK-Y-----ESKRWV-
Y.lipolytica	AWTEEQLA-SMMK-WG-NTRCNM-FWEA-----KLKP-----GHV-PD-DN-KIENFIRTK-Y-----DMKKWA-
D.hansenii	AWTDEQVE-LMIK-WG-NEKCNI-YWES-----KLPD-----GYV-PD-QL-KIDNFIRTK-Y-----DLKKWV-
K.lactis	TWTEEEHLE-AVLE-FGNNKKFNE-YYEN-----KLGG-----GTYVPD-QS-KIGQFIRTK-Y-----ELKKWV-
A.gossypii	TWKEEHLQ-QVVR-FGNNQQANK-VFEG-----RLGG-----GSYVPD-QS-KMGQFIKTK-Y-----EVRKWY-
C.glabrata	TWKEEHLK-ELVQ-MRNNVNANR-VYEA-----KLDPSSKFNGKSLGNDIN-LLQEFIRQK-Y-----ERKRWM-
S.cerevisiae	TWKEEHLV-KLIQ-FKNNLRANS-YYEATL-ADELKQ-----RKI-TD-TS-SLQNFIKNK-Y-----EYKKWI-

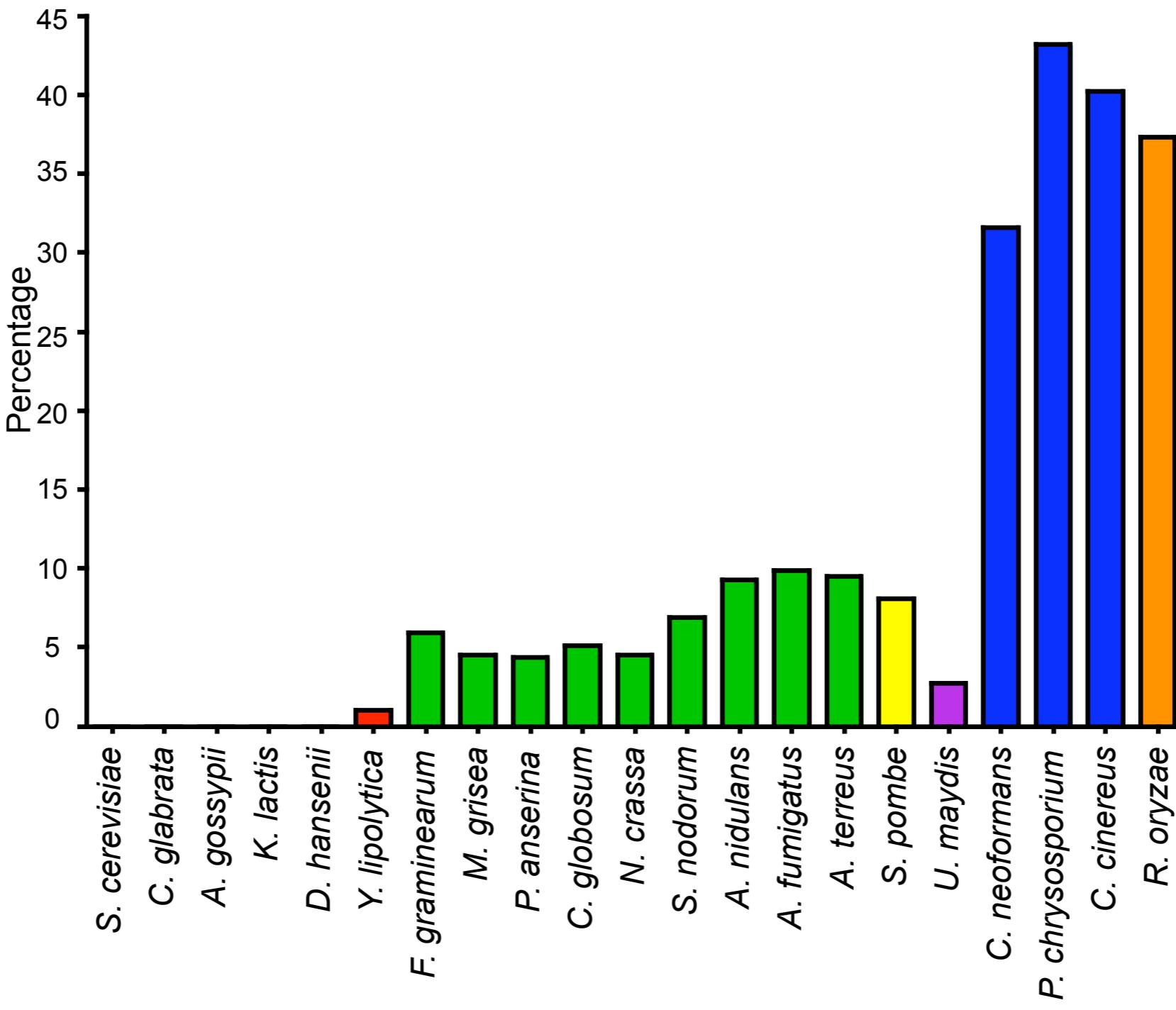
# Patterns of conservation

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# Intron positions shared with animals or plants

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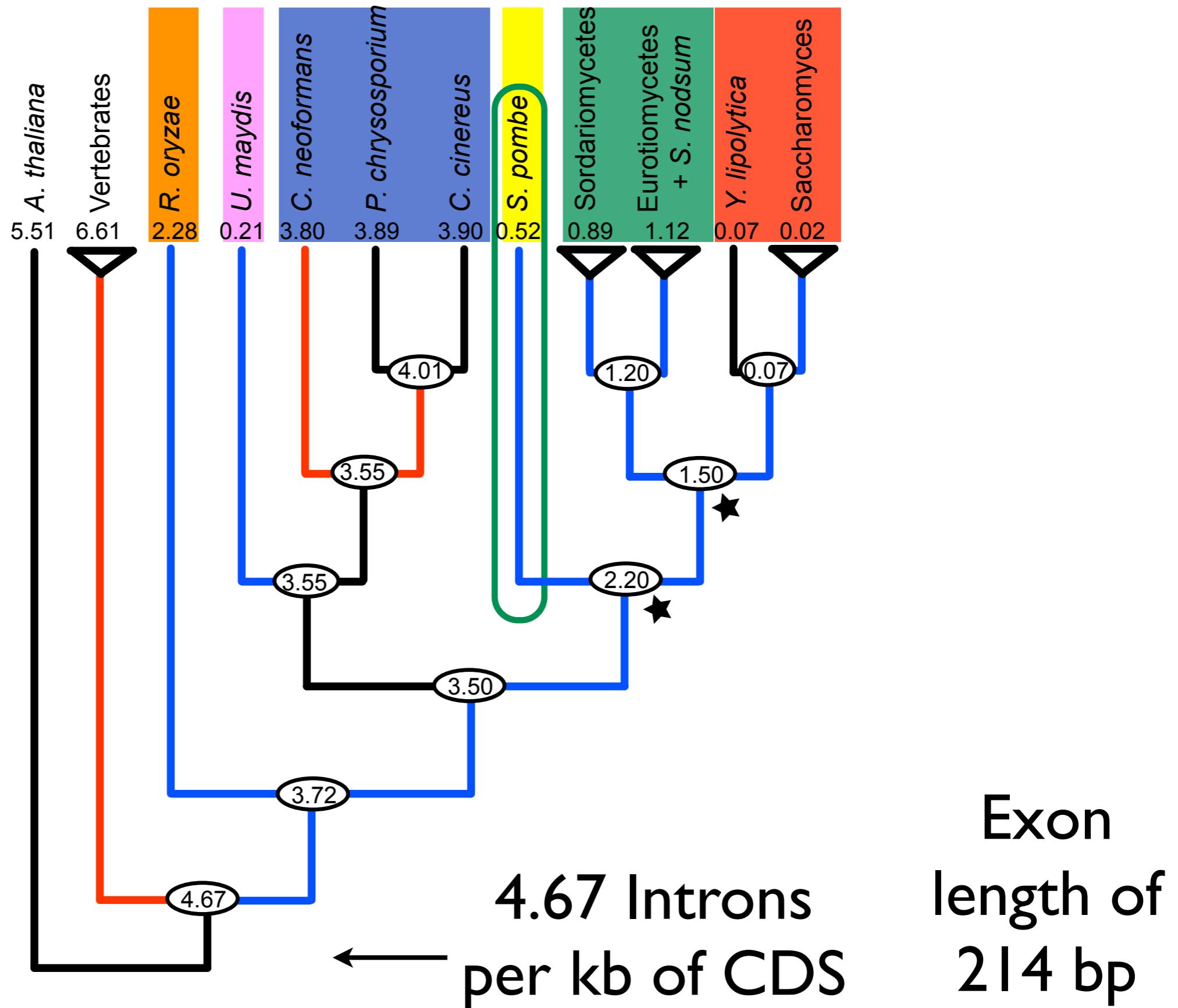


# Intron position reconstruction

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- 3 Methods
  - Roy and Gilbert. 2005
  - Csűrös. 2005
  - Nguyen et al. 2006
- Methods agree for all but 2 nodes in tree

# Reconstruction of ancestral intron densities

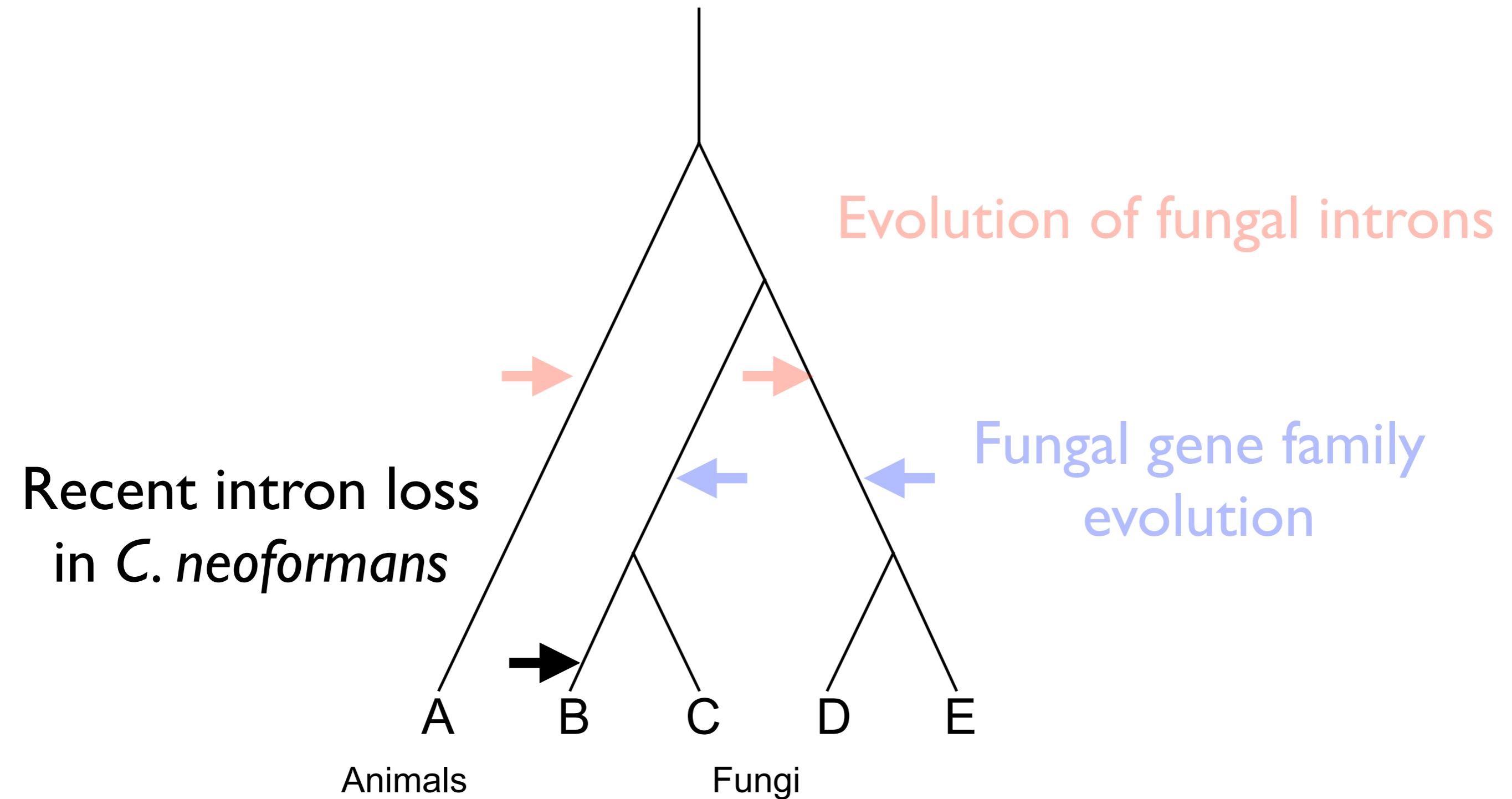


# Conclusions

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- Early eukaryotic crown genes were complex!
  - Ancestor had 70% of the introns in vertebrates - many more than previously reported
- Intron loss has dominated among the fungi
  - **Hemiascomycota** experienced loss
- No significant evidence for intron sliding or double insertions
- Sampling can bias interpretations - all fungi are not equal.

# Fungal comparative genomics



# Mechanism of intron loss in fungi

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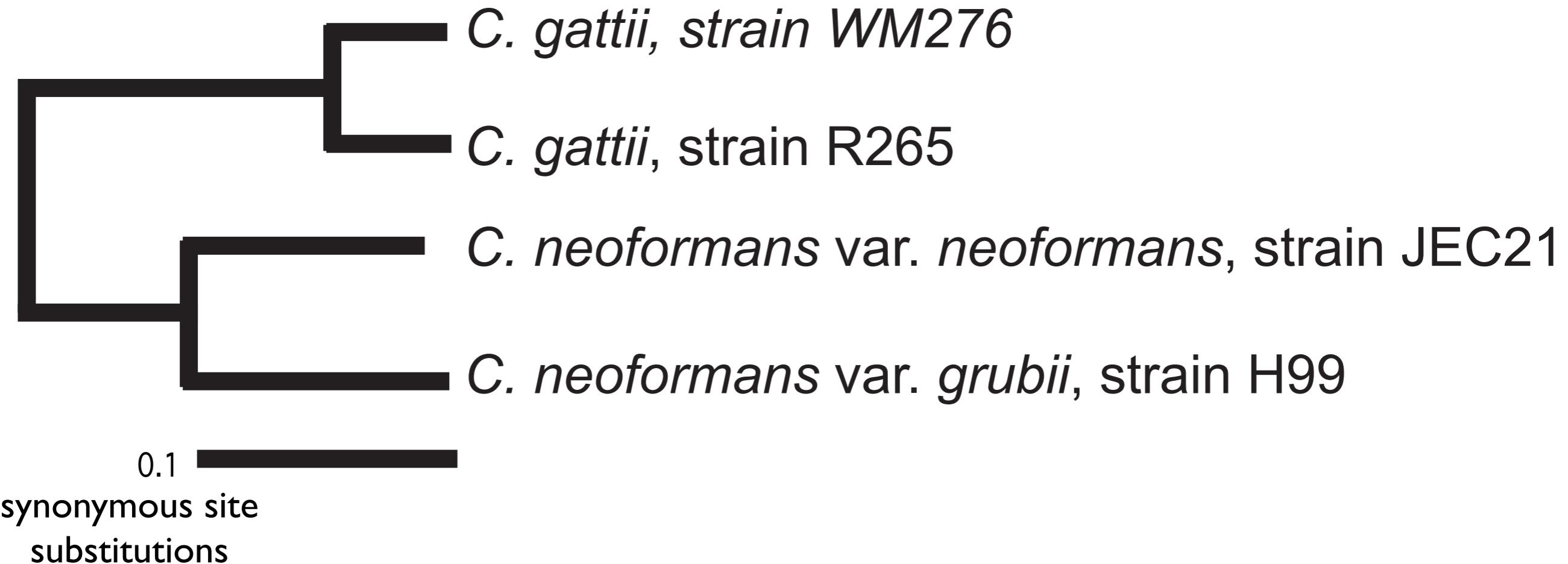
- *S. cerevisiae* and **Hemiascomycota** have undergone intron loss.
- How are introns lost from the genome?
  - Are they lost independently?
  - Are they lost many at a time?
  - What is the molecular mechanism of loss?

# Models of intron loss

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- All introns in *S. cerevisiae* are in 5' end of gene.
- G. Fink proposed transcripts recombine with genome 3' -> 5' explaining 5' retention bias.
- Most lost events in *S. cerevisiae* occurred too long ago to find evidence of mechanism with a comparative approach.

# Sequenced *Cryptococcus* genomes

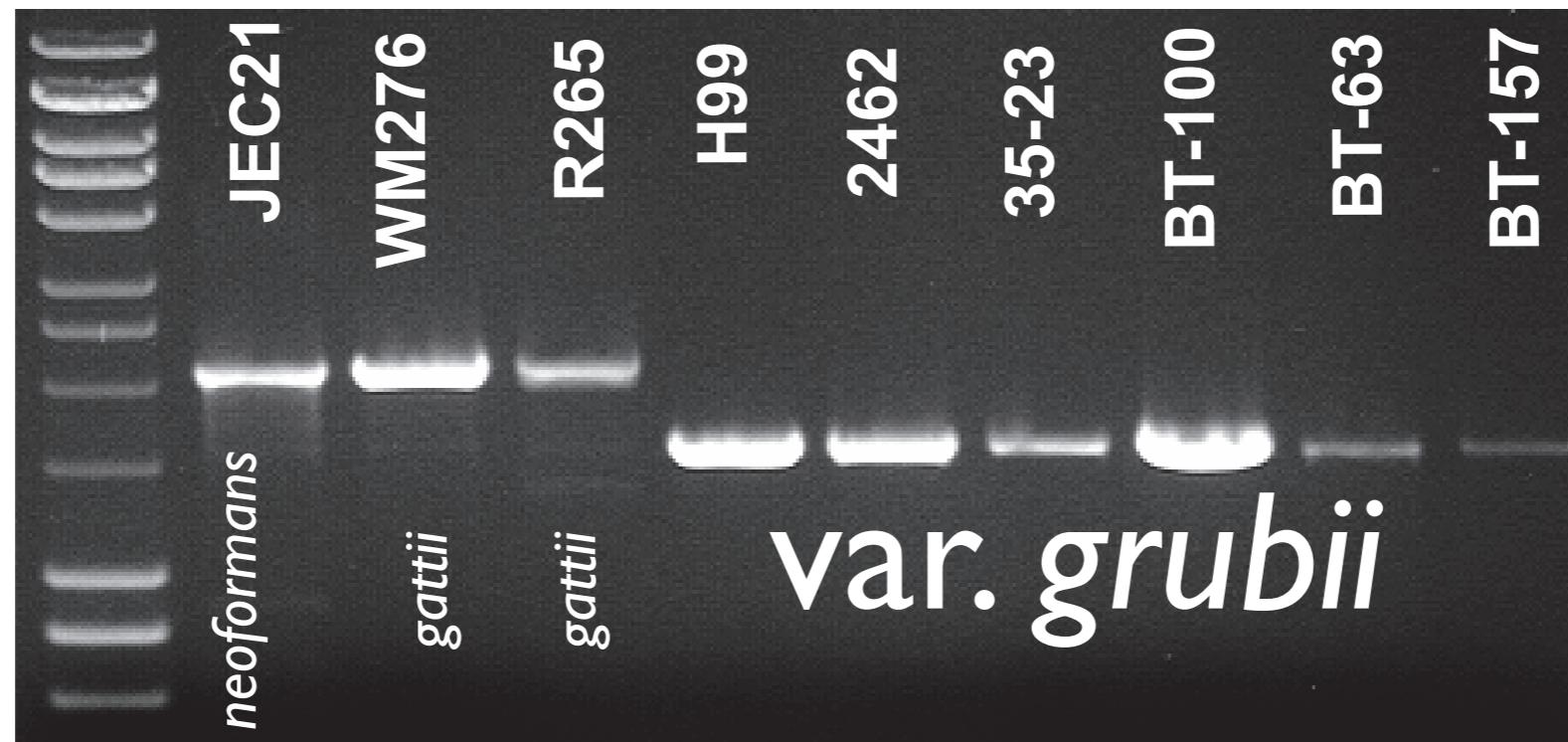


# Screen for intron changes

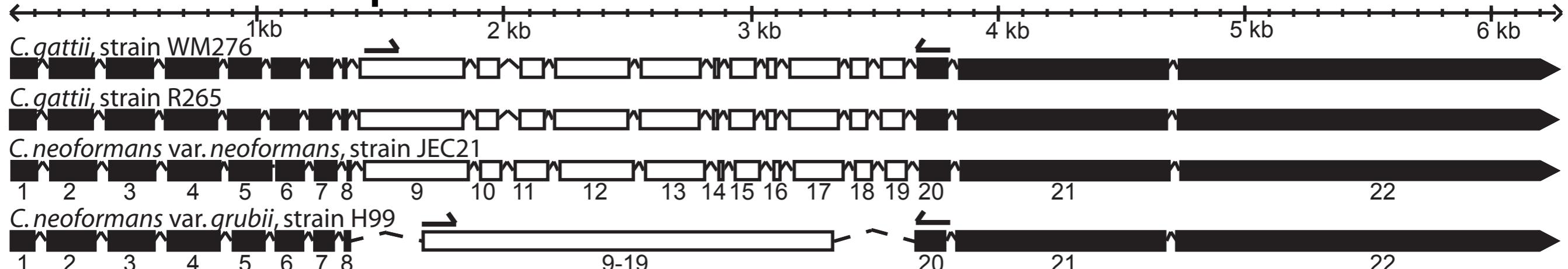
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- Annotate 3 *Cryptococcus* genomes (var. *grubii* and 2 var. *gattii* genomes)
- Identify and align 4-way orthologous genes
  - 5298 orthologous genes (out of ~6500)
- Identify intron position changes

# Intron loss in var. *grubii*



## CNI01550 - putative RNA helicase

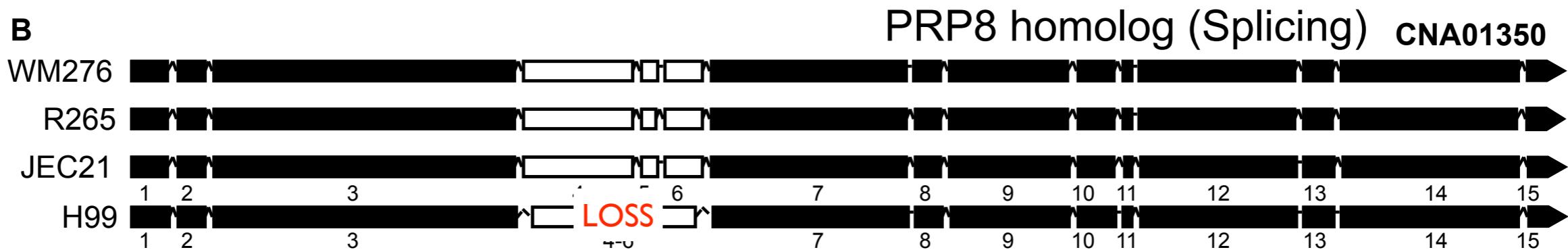
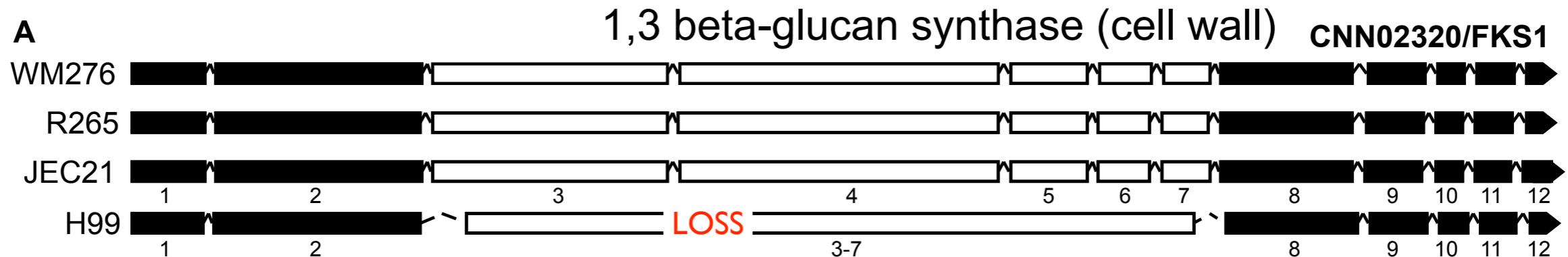


# Intron loss was a precise excision

R265 CGACAAGTACATAAAACTTTTGCTGGCGCAAAGACTTCCATTGCTGACAGAAAACAGGTTGA  
WM276 AGACAAGTACATAAAACTTTTGCTCTCCTCCAAACATTTTTCAATTGCTGACAGAAAACAGGTTGA  
H99 AGACAA ←————— Intron Missing —————→ - GTTGA  
JEC21\_CDS AGACAA-----  
JEC21 AGACAAGTACATACTAGTCCTTGTG---CTATCCAAAGACTTT-CATTGCTGACAGAAAACAGGTTGA  
\*\*\*\*\* \*\*\*\*\*

R265 CGCTGCCGAATTATGTCGATGTTGGAGATTCTTGAGGTAAGCAACAGACTCGAACAGCTTGTTCGGT  
WM276 CGCTGCCGAACTATGTCGATGTTGGAGATTCTTGAGGTAAGCAACAGACTCGAACAGCTTGTTCGGT  
H99 CCCTGCCGAATTATGTCGACGTTGGAGATTCTTGAG-----  
JEC21\_CDS CCCTGCCGAATTATGTCGATGTTGGAGATTCTTGAG-----  
JEC21 CCCTGCCGAATTATGTCGATGTTGGAGATTCTTGAG**GTACGTCGAAACTCGAACAGCTTGTTCGATC**  
\* \*\*\*\*\* \* \*\*\*\*\* \* \*\*\*\*\*

# Other examples of loss



# Conclusions

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- Intron loss via homologous recombination with spliced transcript.
  - Multiple adjacent introns are lost.
  - Precise deletions of introns.
- Loss biased towards the middle of gene not 3'.

# Overall conclusions

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- Multiple genome sequences have helped resolve several outstanding questions in evolution introns
  - Origin of introns still mystery, but early eukaryotic genes were complex
  - Suggested intron function in splicing
- Gene family expansions can be important in identifying molecular basis for adaptation